Safety Assessment of

Microbial Polysaccharide Gums

as Used in Cosmetics

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ABSTRACT

The CIR Expert Panel assessed the safety of 34 microbial polysaccharide gums for use in cosmetics, finding that these ingredients are safe in cosmetic formulations in the present practices of use and concentration. The microbial polysaccharide gums named in this report have a variety of reported functions in cosmetics, including emulsion stabilizer, film former, binder, viscosity increasing agent, and skin conditioning agent. The Panel reviewed available animal and clinical data in making its determination of safety.

INTRODUCTION

This assessment is a review of information relevant to the safety of 34 microbial polysaccharide gums for use in cosmetic formulations. Reported functions for these ingredients include emulsion stabilizer, film former, binder, viscosity increasing agent, and skin conditioning agent.

The Cosmetic Ingredient Review (CIR) Expert Panel has reviewed other non-microbial gums and polysaccharides. In 2012, CIR concluded the Galactomannans, a group of 16 legume polysaccharides, are safe as used in cosmetics.[1] In 2009, the CIR Expert Panel reviewed the safety of Hyaluronic Acid, an amine-derived exopolysaccharide, finding it safe as used.[2] In 1987, the Expert Panel reviewed the safety of Tragacanth Gum (now named Astragalus Gummifer Gum), and concluded that Tragacanth Gum was safe as used; the Panel reaffirmed that conclusion in 2006.[3] Other plant-derived gums that the Panel has reviewed include Acacia Catechu Gum, Acacia Farnesiana Gum, Acacia Senegal Gum; in 2005, the Panel concluded that Acacia Senegal Gum is safe as used, but that the data are insufficient to support the safety of Acacia Catechu Gum and Acacia Farnesiana Gum as used in cosmetics.[4]

Three ingredients included in this safety assessment are hydroxypropyltrimonium chloride compounds. Although hydroxypropyltrimonium chloride itself is not a cosmetic ingredient, it is structurally analogous to some of the compounds included in the CIR safety assessment on trimoniums, in which the CIR concluded that the trimoniums are safe as used in cosmetics when formulated to be non-irritating.[5]

The 34 microbially-produced polysaccharide gums included in this review are:

Xanthan Gum	Sodium Carboxymethyl Dextran
Hydroxypropyl Xanthan Gum	Dextran Sulfate
Undecylenoyl Xanthan Gum	Sodium Dextran Sulfate
Dehydroxanthan Gum	Sclerotium Gum
Xanthan Gum Crosspolymer	Hydrolyzed Sclerotium Gum
Xanthan Hydroxypropyltrimonium Chloride	Beta-Glucan
Gellan Gum	Beta-Glucan Hydroxypropyltrimonium Chloride
Welan Gum	Beta-Glucan Palmitate
Biosaccharide Gum-1	Hydrolyzed Beta-Glucan
Biosaccharide Gum-2	Oxidized Beta-Glucan
Biosaccharide Gum-3	Sodium Carboxymethyl Beta-Glucan
Biosaccharide Gum-4	Pullulan
Biosaccharide Gum-5	Myristoyl Pullulan
Pseudoalteromonas Exopolysaccharides	Levan
Dextran	Rhizobian Gum
Carboxymethyl Dextran	Hydrolyzed Rhizobian Gum
Dextran Hydroxypropyltrimonium Chloride	Alcaligenes Polysaccharides

Some of the polysaccharide gums discussed in this safety assessment can be produced by more than one organism, and sometimes by plants. For example, beta-glucans are produced by fungi, yeasts, and grains[6] and levan can be produced by bacteria, yeasts, or fungi.[7]

Many studies have been conducted with some of the microbial polysaccharide gums in regard to health claims, immunomodulatory activity, anti-oxidant activity, etc. This safety assessment includes only studies and study-types that relate directly to the safety of the cosmetic use of these ingredients.

CHEMISTRY

Definition and Structure

Microbial polysaccharide gums are high molecular weight (MW) carbohydrate polymers that make up a substantial component of the cellular polymers found in and surrounding most microbial cells.[8] These polysaccharide gums are produced by a wide variety of microorganisms and are water soluble gums that have novel and unique physical properties. Microbial polysaccharide gums are generally divided into three groups: exocellular, cell wall, and intercellular.[9] The exocellular polysaccharide gums are those that

constantly diffuse into the cell culture medium and are easily isolated. The cell wall (i.e., structural) and intercellular polysaccharide gums are integral parts of the cell wall or capsular products, and are more difficult to separate from cell biomass.

Microbial polysaccharide gums may be ionic or non-ionic and are primarily linear polysaccharides to which side-chains of varying length and complexity are attached at regular intervals.[8] Most microbial polysaccharide gums are linear hetero-polysaccharides consisting of three to seven different monosaccharides arranged in groups of 10 or less to form repeating units. The monosaccharides may be pentoses, hexoses, amino sugars, or uronic acids. For example, xanthan gum is a polysaccharide produced by a pure-culture fermentation of a carbohydrate with *Xanthomonas campestris*, and is composed of glucose, glucuronic acid, 6-acetylmannose, and 4,6-pyruvylated mannose residues, as seen in Figures 1 and 2.
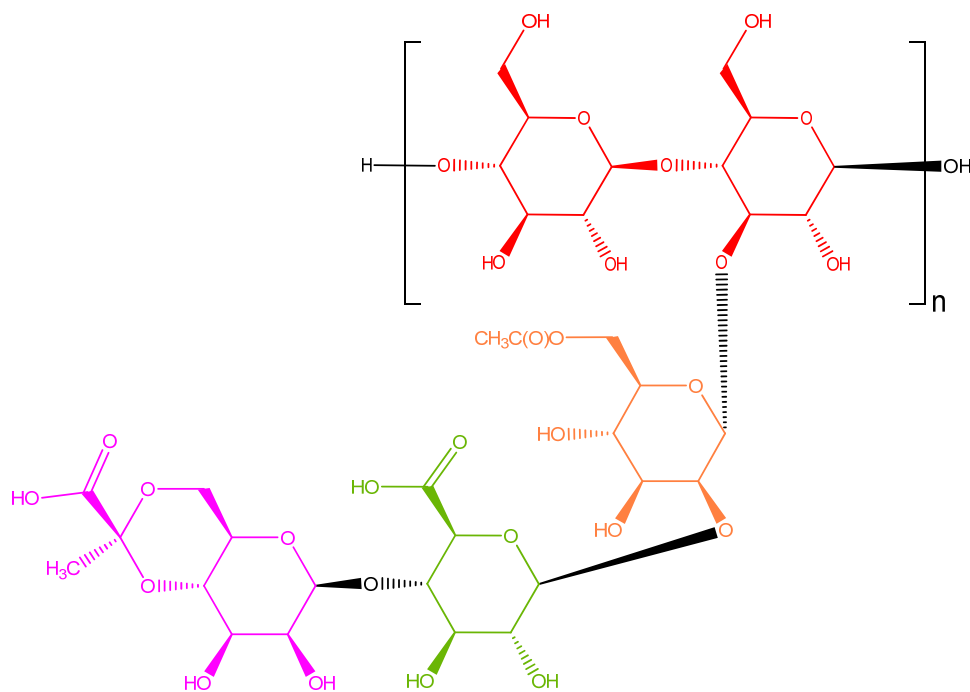

Figure 1. Xanthan Gum – a polysaccharide composed of glucose, glucuronic acid, 6-acetylmannose, and 4,6-pyruvylated mannose
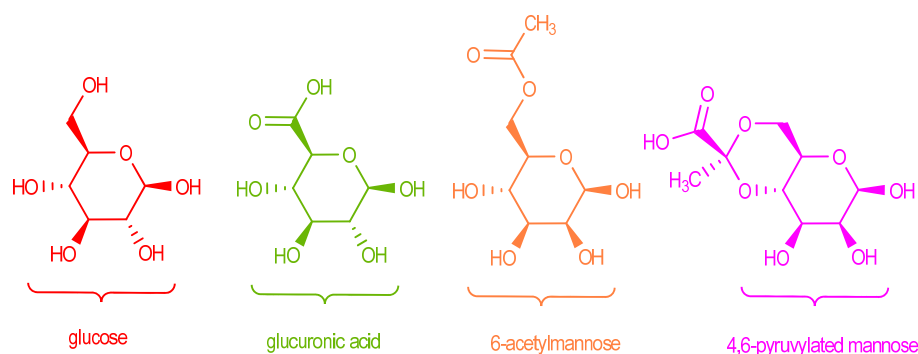

Figure 2. Glucose, glucuronic acid, 6-acetylmannose, and 4,6-pyruvylated mannose, the monosaccharide components of Xanthan Gum.

The other ingredients in this report are related by having similar polymeric repeat units. The definitions and polymeric repeat units of the ingredients included in this review are provided in Table 1.

Physical and Chemical Properties

Available physical and chemical properties are provided in Table 2. The properties of the microbial polysaccharide gums can vary widely based on, among other parameters, the side groups, the ester substituents, or the bacterial strains, but generally speaking, these are very large MW polymers.[10-13]

Constituents/Impurities

The available constituent and impurity data are provided in Table 3.

Method of Manufacture

Methods of manufacture for many of the microbial polysaccharide gums are provided in Table 4. Some of the polysaccharide gums discussed in this safety assessment can be produced by more than one organism, and in some cases, by plants. For example, beta-glucans are produced by fungi, yeasts, and grains[6] and levan can be produced by bacteria, yeasts, or fungi.[7]

USE

Cosmetic

The microbial polysaccharide gums named in this report have a variety of reported functions in cosmetics that include emulsion stabilizer, film former, binder, viscosity increasing agent, and skin conditioning agent.[14] The Food and Drug Administration (FDA) collects information from manufacturers on the use of individual ingredients in cosmetics as a function of cosmetic product category in its Voluntary Cosmetic Registration Program (VCRP). VCRP data obtained from the FDA in 2012,[15] and data received in response to a survey of the maximum reported use concentration by category conducted by the Personal Care Products Council (Council),[16,17] indicate that 19 of the 34 microbial polysaccharide gums named in this safety assessment are currently used in cosmetic formulations. Xanthan gum is used in almost every category of cosmetic product, with 3470 reported uses. Biosaccharide gum-1, sclerotium gum, and beta-glucan are reported to be used in 346, 193, and 137 cosmetic formulations, respectively. All other in-use ingredients have less than 70 uses. The ingredient with the highest concentration of use is pullulan; it is used at up to 12% in leave-on formulations (i.e. tonics, dressings, and other hair grooming aids) and 17% in "other" oral hygiene products (a breath freshener that dissolved in the mouth[18]). Both xanthan gum and biosaccharide gum-1 are used at up to 6% in leave-on formulations and xanthan gum crosspolymer and biosaccharide gum-4 are used at 5% in leave-on formulations. All other in-use ingredients are used at concentrations of ≤3%.

In some cases, reports of uses were received in the VCRP, but no concentration of use is available. For example, sodium carboxymethyl dextran is reported to be used in 10 formulations, but no use concentration data were available. In other cases, no reported uses were received in the VCRP, but a use concentration was provided in the industry survey. For example, hydrolyzed sclerotium gum was not reported in the VCRP to be in use, but the industry survey indicated that it is used in leave-on formulations at up to 1%. It should be presumed that hydrolyzed sclerotium gum is used in at least one cosmetic formulation.

Frequency and concentration of use data are provided in Table 5. The ingredients not listed in the VCRP or by the Council as being used are listed in Table 6.

Products containing some of the microbial polysaccharide gums are reported to be used on baby skin, to be applied to the eye area or mucous membranes, or could possibly be ingested. Some of these ingredients are reported to be used in product types that may be inhaled; for example, dehydroxanthan gum is used in a face and neck spray at 0.2%. In practice, 95% to 99% of the particles released from cosmetic sprays have aerodynamic equivalent diameters in the 10 to 110 μm range.[19,20] Therefore, most particles incidentally inhaled from these sprays are deposited in the nasopharyngeal region and are not respirable.[21,22] Xanthan gum is reported to be used in deodorants at up to 0.6%, and it is not known whether or not these products are sprayed. There is some evidence indicating that deodorant spray products can release substantially larger fractions of particulates having aerodynamic diameters in the range considered to be respirable.[22] However, the information is not sufficient to determine whether significantly greater lung exposures result from the use of deodorant sprays, compared to other cosmetic sprays.

All of the microbial polysaccharide gums named in the report are listed in the European Union inventory of cosmetic ingredients.[23]

Non-Cosmetic

Non-cosmetic uses are of microbial polysaccharide gums are summarized in Table 7. Some of the food and medical use information are described in the following paragraphs.

Xanthan gum[24] and gellan gum[25] are approved as direct food additives in gums, chewing gum bases, and related substances. Xanthan gum is also approved as an indirect food additive.[26] Beta-glucan (as curdlan, a specific beta-glucan that is a linear polymer consisting of β-$(1\rightarrow3)$-linked glucoside residues) is approved as a direct food additive for multipurpose addition.[27] Dextran is an indirect food additive that is generally recognized as safe (GRAS).[28] The World Health Organization (WHO) concluded that studies on the safety of xanthan gum,[29] gellan gum,[30] and pullulan[31] provided sufficient information to be allocated an acceptable daily intake (ADI) of "not specified". Pullulan appears in the Japanese List of Existing Food Additives.[32]

Xanthan gum is used as a stabilizer, thickener, and emulsifying agent in water-based pharmaceutical preparations.[33] Dextran, as dextran 70, is an approved active ingredient for over-the-counter (OTC) use as an ophthalmic demulcent at 0.1% when used with another approved polymeric demulcent.[34] Dextran is used as a plasma volume expander (as dextran 70) and as a blood flow adjuvant (as dextran 40).[35] Sodium dextran sulfate is used as a clinical reagent.

TOXICOKINETICS

Absorption, Distribution, Metabolism, and Excretion

Dermal

In Vitro

Beta-Glucan

A single application of 5 mg/cm^2 of a 0.5% (oat) beta-glucan solution was applied to human abdominal skin.[36] Beta-glucan penetrated the skin into the epidermis and dermis. (No details were provided.)

Human

Dextran

Fluorescent dextrans (FDs) in aqueous (aq.) solution were used to determine the dermal absorption of different MW dextrans (MW 3000 - 70,000, identified as FD-3 and FD-70, respectively) through skin that has been subjected to mini-erosion.[37] Prior to testing, absorption of FD-20 from a test "Cellpatch" (cell) into systemic circulation via a mini-erosion site was verified in two male subjects and one female subject. The largest of the test molecules filtered from the blood into the urine was 20,000 MW, and with increasing erosion diameter, absorption increasingly occurs via the lymphatic system. Each subject received a single cell containing 0.5 ml solution (5 mmol). After 24 h, a mean of 54.3% of the total FD-20 dose had been absorbed, and 12.6% of the absorbed dose was recovered in the urine.

The effect of molecular size on absorption of FDs was then determined. Four cells with 100 μl of FD-3, FD-10, FD-20, or FD-70 in 0.5 ml isotonic saline were applied for 24 h to 6 mm mini-eroded sites on 4 male and 3 female subjects. A fifth cell, without FD, was applied as a negative control. The FD concentration in each cell was measured using spectrofluorometry at various times. The dextrans were readily absorbed, but absorption decreased with increasing molecular size. The absorption of FD-3 was 37.9%, but for FD-70, it was 20.1%. Further testing using 3 male and 3 female subjects determined that the degree of transdermal absorption was directly related to the area of erosion; 20.5% of FD-3 absorbed through a 3 mm erosion area, while 60.7% of the same molecular size FD absorbed through a 10 mm erosion.

Oral

Non-Human

Xanthan Gum

Rats were fed a diet containing 2% [^{14}C]xanthan gum that was produced by fermentation of uniformly-labeled glucose with *Xanthomonas campestris*.[38] No accumulation was found in the tissues. A maximum of 15% of the radioactivity was metabolized to carbon dioxide within 100 h. Fecal analysis indicated that there was no accumulation of the polysaccharide material, except acetate. (Acetate and pyruvate accounted for only 9.8% of the label in the gum used). In the feces, 98% of the radioactivity was attributed to unchanged or slightly modified polysaccharide. In vitro testing indicated that non-enzymatic hydrolysis and fecal microorganisms were responsible for the in vivo breakdown of xanthan gum. (No additional details were provided.)

Gellan Gum

In animal feeding studies using radiolabeled gellan gum, the majority of the gellan gum that was administered was recovered in fecal matter.[13] This appears to indicate that no endogenous enzymes that are able to break down gellan gum are present in the small intestine. (No additional details were provided.)

The absorption, distribution, and excretion of gellan gum was determined in studies using a dually-radiolabeled gum that was prepared in separate fermentations using [^{3}H]glucose and [^{14}C]glucose as carbon sources.[39] (The ^{3}H product was subjected to multistage purifications for a relatively pure [^{3}H]polysaccharide, which was then added to the ^{14}C fermentation, giving a polysaccharide fraction that was dual-labeled and a non-polysaccharide fraction labeled only with ^{14}CO$_2$.) In the first study, one male and one female Sprague-Dawley rat were dosed by gavage with a single dose of 960 mg/kg [^{3}H/^{14}C]gellan gum (4 μCi). Expired air was collected for 24 h after dosing, and <0.55% of the dosed radioactivity was detected as ^{14}C.

Four male and three female Sprague-Dawley rats were then given a single dose by gavage of 870 mg/kg [^{3}H/^{14}C]gellan gum (2.9-4.1 μCi ^{14}C; 0.7-0.9 μCi ^{3}H). Urine and feces were collected for 7 days. Approximately 86% of the dosed ^{14}C was excreted in the feces and 2-3% in the urine, and approximately 100% of the dosed ^{3}H was excreted in the feces and 4% was in the urine. Tissue and carcass ^{14}C radioactivity was approximately 3-4% of the dose. The ^{3}H activities in the tissues were below the limits of accurate quantification.

In the last study, one male and four female Sprague-Dawley rats were dosed with 1 g/kg [^{3}H/^{14}C]gellan gum by gavage, and blood samples were taken at various intervals over a 7-day period. (It is not stated, but it is appears that one dose was administered). The peak level of radioactivity in blood, equivalent to 0.4% of the dose, occurred about 5 h after dosing.

Dextran

Groups of 5 fasted male Sprague-Dawley rats were given a single dose by gavage of 50 mg/kg fluorescein-labeled dextrans (FD-4, 4400 avg. MW; FD-20, 19,000 avg. MW; or FD-40, 40,500 avg. MW).[40] The dextran solution was prepared as 25 mg/ml in iso-tonic phosphate buffer. Blood samples were taken at various intervals for up to 4 h after dosing with FD-4, up to 8 h after dosing with FD-20, and for up to 24 h with FD-40. Urine samples were taken at intervals for up to 8 h after dosing with FD-4 and FD-20 and for up to 24 h after dosing with FD-40. None of the dextrans were detected in the serum after oral administration. Small amounts of the dose, ranging from 0.308% with FD-4 to 0.0138% FD-40, were detected in the urine. The oral bioavailability was 0.398, 0.0728, and 0.0431% for FD-4, FD-20, and FD-40, respectively.

Dextran Sulfate

Two groups of 5 male Wistar rats were dosed by gavage with 5 mg/ml of 20 mg/kg dextran sulfate containing 12 μCi ^{3}H-dextran sulfate/kg (8000 avg. MW), and each group was killed 3 or 24 h after dosing.[41] Most of the radioactivity was detected in the feces; 22.5% of the dose was recovered after 24 h. Metabolites, breakdown products, and ^{3}H$_2$O were not found in the feces, and the re-searchers stated that it was most likely that the dose recovered in the feces was unabsorbed dextran sulfate. Approximately 10% of the dose was recovered in the urine after 24 h, with 6% recovered after 3 h. Urinary ^{3}H elution profiles indicated that intermediate MW metabolites or breakdown products were formed and that either intact or partially intact dextran sulfate was absorbed through the epithelium of the gastrointestinal (GI) tract. The 6-24 h samples indicated a marked shift towards smaller MW products.

Beta-Glucan

Two male Sprague-Dawley rats were dosed orally with 20 mg/kg bw [U-^{14}C]beta-glucan (as curdlan) in water prepared from [U-^{14}C]glucose.[42] Most of the radioactivity was recovered in expired CO_2; 77% of the dose was recovered in 24 h and 89% in 72 h. A total of 7.7 and 12% of the radioactivity as administered dose was recovered in the feces after 24 and 72 h, respectively, and 2.6 and 3.3% was recovered in the urine after 24 and 72 h, respectively.

In another study, three male Wistar rats were dosed orally with 20 mg/kg bw [^{14}C] beta-glucan (as curdlan) in water.[42] Initially (i.e. the first 3 h), the excretion of $^{14}CO_2$ was low, but then increased linearly up to 12 h, plateauing at 39% of the administered radiolabel. A total of 3.4 and 3.8% of the radioactivity as administered dose was recovered in the feces after 24 and 48 h, respec-tively, and 1.3 and 1.4% was recovered in the urine after 24 and 48 h, respectively. In a group of three male Wistar rats, admini-stered 5 mg/ml tetracycline for 5 days prior to and 2 days following beta-glucan, it was demonstrated that the intestinal microflora are partly responsible for the metabolism of beta-glucan to carbon dioxide.

The effect of dose on metabolism was also examined.[42] Three male Wistar rats were given an oral dose of 2.3, 23, or 230 mg/kg bw [^{14}C]beta-glucan (as curdlan) in water. At the two higher doses, excretion of radioactivity as carbon dioxide decreased with in-creasing dose, while fecal excretion of the radiolabel increased. The researchers stated that this was an indication of limited me-tabolism at higher doses.

Pullulan

Five fasted male Wistar rats were dosed by gavage with a 2 ml of a 10% solution of pullulan (49,000 MW) in 0.9% saline.[43,44] The animals were killed 1 h after dosing and the contents of their stomach and small intestines were collected. Approximately 3% of the pullulan had been hydrolyzed; it was not known whether the hydrolysis products were absorbed by the small intestine.

Human

Dextran

Dextran can be depolymerized by α-1-glycosidases (dextranases) that occur in the liver, spleen, kidney, and lower part of the GI tract.[10]

Dextran Sulfate

Six fasted male subjects were given a single oral dose of 1800 mg dextran sulfate (7000-8000 MW; 17-20% sulfur) and, after 48 h, a single intravenous (i.v.) dose of 225 mg dextran sulfate in saline infused over 60 min.[45] After oral dosing, no measurable dextran sulfate was found in the plasma using the competitive binding assay, and there was no increase in activated partial thromboplastin time (APTT). Plasma lipolytic activity did not increase the first 3 h after oral dosing; at 3-4 h after oral dosing, it increased by two times the baseline average. Very little dextran sulfate was recovered in the urine after oral dosing. After i.v. dosing, peak plasma concentrations were 26-35 μg/ml, and the APTT was increased by an average of 6.9 times over the baseline values. The plasma lipolytic activity increased by an avg. of 438 times the baseline value. Dextran sulfate was recovered in the urine after i.v. dosing.

Pullulan

Pullulan is only partially hydrolyzed by salivary and pancreatic amylases of the upper GI tract; essentially no monomeric glucose is released during hydrolysis.[44] Pullulan is largely resistant to digestion in the GI tract because of the occasional presence of 1→3-glycosidic linkages and the high percentage of α-1→6-glycosidic linkages.[31] The degree of digestion appears to be dependent on

molecular mass. Pullulan is fermented in the colon by intestinal microflora to produce short-chain fatty acids; the degree of fermentation is dependent on the degree of polymerization of pullulan.

Six subjects ingested 10 g pullulan (50,000 MW) for 14 days.[44,46] Administered pullulan was fully digested in the intestinal tract and was not detected in the feces. After 14 days, the fecal short chain fatty acid concentration increased from 6 mg/g to 8.8 mg/g. The researchers concluded that pullulan was completely fermented to short-chain fatty acids by intestinal bacteria.

Parenteral
Non-Human
Dextran

Rabbits were given a daily i.v. 30 ml dose of a 6% solution of partly hydrolysed bacterial dextran (75,000 avg MW) 6 days/wk for 103-113 wks.[47] Eleven animals were evaluated. Approximately 25% of the dose was excreted in the urine. The plasma concentration of dextran at study termination (0.50 g/100 ml) did not differ from the value at 2 mos (0.44 g/100 ml), but it was generally greater than the value reported at 24 h after a single 30 ml dose (not given). Moderate dextran storage was observed in the spleen without an increase in the total carbohydrates. However, considerable storage was observed in the liver with a marked increase in carbohydrates. (Additional details and results are provided in the section on 'Repeated Dose Toxicity').

Groups of 5 male Sprague-Dawley rats were given a single i.v. dose of 5 mg/kg FD-4 (4400 avg. MW), FD-20 (19,000 avg. MW), FD-40 (40,500 avg. MW), FD-70 (71,000 avg. MW), or FD-150 (147,800 avg. MW).[40] The dextran solution was prepared as 5 mg/ml in isotonic phosphate buffer. Blood samples were taken at various intervals for up to 4 h after dosing with FD-4, up to 8 h after dosing with FD-20, and for up to 24 h with FD-40, FD-70, and FD-150. Urine samples were taken at intervals for up to 8 h after dosing with FD-4 and FD-20 and for up to 24 h after dosing with FD-40, FD-70, and FD-150. Pharmacokinetic parameters were MW-dependent. Concentrations of the three highest MW dextrans could be detected in serum for up to 12 h after dosing, while FD-20 and FD-4 were not found in the serum after 3 and 1.5 h, respectively. The distribution half-life ($t_{1/2}\alpha$) ranged from 0.0517 to 0.895 h for FD-4 and FD-150, respectively, and the elimination half-life ($t_{1/2}\beta$) ranged from 0.282-3.03 h for FD-4 and FD-150, respectively.

Male Sprague-Dawley rats were also given a single i.v. dose of 1, 25, or 100 mg/kg FD-4 (avg. MW 4300) and FD-150 (avg. MW 145,000).[48] The dextran solution was prepared as isotonic phosphate buffer at a concentration that would result in a test volume of 2 ml/kg. Groups of 4 rats each were used for blood, urine, and tissue samples at various intervals for up to 6 h in rats dosed with FD-4 and for up to 96 h in rats dosed with FD-150. Renal excretion was a major excretion pathway for FD-4 but not FD-150. Urinary recovery ranged from 79-82% of the dose with FD-4 and only from 1.1-2.1% of the dose with FD-150; at each MW, the dose administered did not have a statistically significant effect on the amount excreted. Renal clearance ranged from 344-360 ml/h/kg with FD-4 and from 0.131-0.245 ml/h/kg for FD-150, and systemic clearance ranged from 420-457 ml/h/kg for FD-4 and from 8-20 ml/k/kg for FD-150. The highest concentrations of FD-4 were found in the kidneys at 1 min after dosing (9.31%, 10.0%, and 10.4% of the dose with 1, 25, and 100 mg/kg, respectively) was linear with dose. The highest concentrations of FD-150 were found in the liver (68.5% at 5 h, 51.6% at 24 h, and 41.5% of the dose at 24 h with 1, 25, and 100 mg/kg, respectively) and the spleen (11.5% at 12 h, 2.09% at 48 h, and 1.21% of the dose at 96 h with 1, 25, and 100 mg/kg, respectively); recovery of dextran in the liver and spleen was non-linear, with a greater difference seen in the spleen than in the liver. The researchers reported that the MW of recovered FD-4 remained relatively constant, but that of recovered FD-150 changed significantly. The MW of FD-150 recovered in the urine was <40,000, and, in the liver, the avg. MW at 96 h was ~70,000. The estimated MW of the recovered FD-150 in the liver appeared to be dose-dependent with higher doses having a higher avg. MW. The researchers concluded that excretion of lower MW dextrans was independent of dose, while excretion of higher MW dextrans was dose-dependent.

Another study also found that MW affected the distribution and excretion of dextran.[49] Female BALB/cCrSlc mice were dosed i.v. with 0.1 ml of 0.1 wt% ^{125}I-labeled dextran, MW ranging from 4980-220,000, in phosphate buffered saline (PBS); dextran was labeled through radioiodination of tyramine residues. Blood samples were taken at various intervals for some groups, and tissues were collected from others. (The number of animals/group was not specified.) High MW dextran remained in the blood longer than lower MW dextran. The $t_{1/2}\beta$ also increased with increasing MW, with a pronounced change occurring around 30,000 MW. After 3 h, 83% of the dose of the lowest MW dextran was excreted, while only 41% of the highest MW dextran was found in excrement. Most of the high MW dextran was found in the liver; after 3 h, 5.2% of the high MW dextran was found in the liver, while only 0.7% of the lowest MW was recovered in that tissue. At 3 h, 3.5 vs. 19% of the dose of the lowest and highest MW dextran, respectively, was recovered in the carcass. Two to 10% of the dose was recovered in the GI tract, but the amount recovered did not appear to be related to MW.

Dextran Sulfate

A preliminary study was performed in which 2 male Wistar rats were dosed by i.v. injection with 20 mg/kg dextran sulfate 8000 avg. MW) containing 10 μCi [³H]dextran sulfate/kg via the penile vein (5 mg/100 μl).[41] The rats were killed 1 or 3 h after dosing. The total ³H excreted in the urine accounted for approximately 50% of the administered dose. Within 1 h after i.v. administration, rapid excretion of intact dextran sulfate occurred.

In the main study, groups of 5 male Wistar rats were dosed by i.v. injection via the penile vein with 5 mg/100 μl of 20 mg/kg dextran sulfate containing 12 μCi [³H]dextran sulfate/kg, and each group was killed 3 or 24 h after dosing. Urine was the major route of excretion following i.v. dosing, with approximately 46% of the dose excreted within 3 h and 51% within 24 h. Approximately 2% of the dose was recovered in the feces after 24 h. Based on the ³H elution profiles, it was hypothesized that dextran sulfate or its metabolites were incorporated into higher MW compounds, such as glycogen, at 3-6 h and smaller MW at 6-24 h. In the plasma, the highest concentration of dextran sulfate was found at 3 h; the amount decreased with time.

In the tissues, the highest amounts of radioactivity were distributed in the liver, kidney, and spleen. The amount of radioactivity recovered in the tissues following i.v. administration was compared to that found following oral administration. (The oral study was described previously in this safety assessment.) Concentrations of ³H in all tissues and fluids were statistically significantly higher in the animals dosed by i.v. injection compared to those in animals dosed orally.

Beta-Glucan

Groups of two male Sprague-Dawley rats were dosed by intraperitoneal (i.p.) injections with 20 mg/kg bw [¹⁴C]beta-glucan (as curdlan) in water, and the animals were killed 0.5, 3, 6, or 24 h after dosing.[42] After 24 and 48 h, only 1.8% and 4.1% of the radioactivity was recovered in CO_2, respectively, 0.05 and 0.12% was recovered in the feces, respectively, and 3.5 and 4.1% of the radioactivity as % of dose was recovered in the urine, respectively. Whole-body radioautography showed that the radiolabel was distributed in the intestinal fluids.

Pullulan

The effect of MW on the distribution and excretion of pullulan was examined.[49] Female BALB/cCrSlc mice were dosed i.v. with 0.1 ml of 0.1 wt% [¹²⁵I]pullulan, MW ranging from 5800-853,000, in PBS; pullulan was labeled through radioiodination of tyramine residues. Blood samples were taken at various intervals for some groups, and tissues were collected from others. (The number of animals/group was not specified.) High MW pullulan remained in the blood longer than lower MW pullulan. The $t_{1/2}\beta$ also increased with increasing MW. After 3 h, 96% of the dose of the lowest MW pullulan was excreted, while only 15% of the highest MW pullulan was found in excrement. At 3h, most of the high MW pullulan, 50% of the dose, was recovered in the liver; only 1% of the dose of the lowest MW pullulan was recovered in the liver after 3 h. The highest percentage of the dose recovered in the GI tract and the carcass was found with mid-level MW pullulan; 5.3% of the dose was recovered in the GI tract with 100,000 MW pullulan and 10.1% of the dose was recovered in the carcass with 48,000 MW pullulan.

Fasted male Wistar rats (number not specified) were injected with a single i.v. dose 2 ml of 0, 6, 12, 18, or 24 mg/kg fluorescein-labeled pullulan (MW 58,200) in saline in the jugular vein.[50] Pullulan was rapidly eliminated from the blood; however, elimination decreased as dose increased. Hepatic uptake was also dose-dependent; the hepatic uptake clearance of pullulan decreased with increased dose. In the liver, distribution of pullulan in the parenchymal cells was 2.5 times greater than in the nonparenchymal cells. The researchers did state that with a higher MW pullulan, avg. 70,000 MW, uptake was greater in the nonparenchymal cells.

Human
Dextran

Four male and three female subjects received an i.v. injection 100 ml a partially degraded dextran (40,000 avg. MW; 10% w/v in 0.9% saline).[51] Blood samples were taken at various intervals after dosing. There was an initial rapid decrease in the serum in the first hour after dosing. Different fractions of the dextran were eliminated from the plasma at different rates; higher MW fractions remained in the plasma longer.

TOXICOLOGICAL STUDIES

Single Dose (Acute) Toxicity

Acute toxicity studies are summarized in Table 8. The acute toxicity of xanthan gum, gellan gum, beta-glucan, sodium carboxymethyl beta-glucan, and pullulan was assessed orally in mice, rats, and/or dogs, and dextran sulfate and beta-glucan were tested by i.p. and i.v. dosing in mice and rats. There was no notable toxicity observed in these studies. In acute inhalation studies, the LC_{50} of xanthan gum was >21 mg/l in rabbits and of gellan gum was >5.06 mg/l in rats. A single 180-min exposure of humans office dust containing 10 mg curdlan/g dust resulted in decreased nasal volume, in swelling in the nasal turbinates, and an increase in nasal eosinophils when compared to "clean" dust.

Inflammatory Response
Beta-Glucan

The inflammatory response following a single exposure to beta-glucan (as curdlan) was evaluated in guinea pigs.[52-54] Mostly, no effect or a slight decrease in inflammatory cells in lung lavage was observed. A 4 h inhalation exposure to 1% beta-glucan in dust (mass mean aerodynamic diameter [MMAD] 5 μm) by guinea pigs produced a delayed subacute nasal congestion when compared to dust without beta-glucan (MMAD 6.5 μm) and resulted in decreased nasal volume; after 18 h, there was a significant decrease

in nasal volume.[52] In humans, inhalation exposure to beta-glucan in dust for four 3 h exposures resulted in increased nasal swelling and decreased nasal volume and an immediate increased in nasal eosinophil/ml count.[55]

Pullulan

The effect of pullulan on inflammation was investigated in ICR mice using the xylene-induced acute inflammatory mouse ear model.[56] Groups of nine mice were dosed orally with 0, 62.5, 125, or 250 mg/kg pullulan in distilled water 30 min prior to topical application of xylene to one ear. Two h after xylene application, all animals were killed. Compared to xylene-treated controls, ear weights were statistically significantly decreased in a dose-dependent manner. Additional histological indicators of inflammation were not observed.

Levan

An interleukin (IL)-1α release assay was used to determine the anti-inflammation effect of a 5% aq. levan solution on artificial skin.[57] Primary skin irritation was first induced using sodium lauryl sulfate (SLS). A dose of 0.01 or 0.05 mg/ml of the solution was applied to the skin. Levan decreased IL-1α release, indicating an anti-inflammatory effect.

Cytotoxicity

Levan

The cytotoxic effect of 5% levan (w/w) was determined using human fibroblasts and keratinocytes.[57] The 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay was used to measure cell viability and proliferation after 24 h incubation with levan. Levan, ≤100 µg/ml, was not cytotoxic to human fibroblasts. Levan had a proliferative effect in keratinocytes; proliferation was >30% at concentrations of >1 mg/ml.

Repeated Dose Toxicity

Repeated dose toxicity studies are summarized in Table 9. The oral toxicity of xanthan gum was evaluated in rats and dogs, of gellan gum in rats, dogs, and monkeys, of dextran in rats, of beta-glucan in mice, rats, and dogs, and of pullulan in rats. Most of the studies were dietary, and study durations lasted up to 2 yrs. Most observations were related to changes in feed consumption and intestinal effects. In guinea pigs, inhalation exposure to 100 µg/ml beta-glucan (as curdlan), 4 h/day, 5 days/wk for 4 wks, did not have an effect on the cells of the lung lavage or cell wall, and there were no microscopic lesions of the lung. With i.p. administration, no toxicity was reported when mice were dose 10 times over 2 wks with 5 mg xanthan gum in 0.5 ml water or mice or guinea pigs were dosed with 250 mg/kg bw beta-glucan for 7 days. Intravenous administration of 40 and 1000 mg/kg bw beta-glucan for 30 days resulted in hepatosplenomegaly in mice.

Oral Intake by Humans

Xanthan Gum

Five male subjects consumed 150 mg/kg bw/day xanthan gum as three measured portions daily for 23 days.[58] Ingestion of xanthan gum had no significant adverse effect on hematology, clinical chemistry, or urinalysis parameters.

Gellan Gum

Five male and five female subjects consumed a daily dose of 175 mg/kg gellan gum for 7 days and 200 mg/kg/day for the next 16 days.[58] No adverse dietary or physiological effects and no allergenic effects were reported.

Dextran and Pullulan

No adverse effects were seen in a study in which 10 g pullulan, dextran, and soluble starch were ingested by 8 male volunteers.[42,59] Each ingredient was administered for 14 days, and there was a 14-day wash-out period between treatments.

Beta-Glucan

Six male subjects ingested milkshakes with or without beta-glucan (as curdlan) for 28 days.[42] The test subjects were given 6 g/day for 5 days, 35 g/day for 2 wks, and 50 g/day from day 21-28. No evidence of toxicity was observed.

Pullulan

In a tolerance study, 13 subjects ingested 10 g/day of pullulan (50,000 MW) for 14 days.[44,46] There were no effects on clinical chemistry parameters, and no adverse effects were reported.

Industrial Exposure

Xanthan Gum

The relationship between the handling of xanthan gum powder and adverse symptoms was examined using exposure groups based on average percentage of time spent in a plant that uses a fermentation process to manufacture xanthan gum, not on the basis of expected intensity of exposure to xanthan gum.[60] The exposure of interest was to the employees exposed to milling, blending, and packaging of the product; a total of 39 employees were surveyed. Analysis of the results found no significant acute or chronic effects in pulmonary function in any of the exposure groups.

REPRODUCTIVE AND DEVELOPMENTAL TOXICITY

Oral

Xanthan Gum

A three-generation reproductive toxicity study was performed in which albino rats were fed dietary levels of 0, 0.25, and 0.5 g/kg bw/day xanthan gum.[61] Ten males and 20 females were used for the first generation, and 20 males and 20 females were used in the next two generations. Rats were mated to produce two litters per generation, and the successive generations were selected from weanlings of the second litter. Survival and reproductive parameters were similar for treated and control parental rats. Body weights of treated parental rats were slightly decreased compared to controls in each generation. There were no significant differences in developmental parameters between test and control litters, and no malformations were observed in any of the offspring.

Gellan Gum

Groups of 25 gravid female Sprague-Dawley rats were fed a diet containing 0, 2.5, 3.8, or 5.0% gellan gum (varied degree of acetylation; 58.5% polysaccharide) on days 6-15 of gestation.[39] No fetotoxic or teratogenic effects were reported. (No other details were provided.)

Beta-Glucan

A developmental study was performed with 0, 5, or 15% beta-glucan (as curdlan) with a control group of 40 male and 80 female CD rats and test groups of 20 male and 40 female rats.[42] The animals, which were mated twice, were fed the test diet throughout the study. Twenty of the treated dams nursed their own litters; the other 20 treated dams switched litters with the control dams so that treated animals would nurse control pups and control animals would nurse test pups. The F_{1a} offspring were killed prior to the second mating.

No changes in mortality, behavior, or appearance were observed. Male sires of the 15% beta-glucan group had decreased growth rates compared to controls, and males and females of the 15% group had decreased feed consumption. At birth, there were no differences in fertility or lactation among the groups, and no abnormalities were reported. However, survival of the F_{1a}, but not the F_{1b}, pups of the 5% group was statistically significantly decreased compared to controls. Weight gain of all F_{1a} litters of treated dams that nursed their own pups was statistically significantly decreased compared to controls; for the F_{1b} litters, the difference was statistically significant only in the 15% group. Statistically significant decreases in weight gain were also observed for pups of treated dams that were nursed by control dams, but the effect was reduced. Statistically significant decreased weight gain at some intervals was also observed for control pups nursed by treated dams. A no-observable effect level (NOEL) was not established.

The researchers also examined whether there would be decreased weight gain by the pups if dosing was discontinued during lactation. The protocol was similar to that just described, except that all groups consisted of 20 male and 40 female CD rats, and there was no cross-over at lactation. Weight gain by all pups during lactations was similar, although the researchers did state that the pups could have consumed parental diet from day 10+. The NOEL for parental toxicity and embryotoxicity was 15% beta-glucan.

A three-generation reproductive study was performed in which groups of 20 male and 40 female CD rats were fed a diet containing 0, 1, 5, or 15% beta-glucan (as curdlan) for 100 days.[42] The F_0 parents were mated twice, and the number of parents was halved after weaning of the first litter. The F_1 parents were mated three times and the F_2 parents were mated twice. The F_{1b} and F_{2b} litters were used to produce the next generation. After the third mating of the F_1 parents, half of the F_1 dams were killed on day 13 of gestation, and the remaining dams were killed on day 20 of gestation.

Mean growth and feed consumption were slightly decreased in male parental rats of the F_0 and F_1 generations of the 15% group. No gross or microscopic changes were observed in F_2 parents. No treatment-related effects on reproductive and developmental parameters were observed, but body weights of pups in almost all litters in all generations were statistically significantly decreased during lactation in the 15% group. Biologically-significant differences in body weights were not seen in litters of the other dose groups. No gross or microscopic lesions were observed in the F_{3b} pups of the 15% group. In the F_1 parents killed after the third mating, no reproductive or developmental effects were observed. Mean fetal weights in all groups were statistically significantly decreased compared to controls; however there was no dose-response. The NOEL for parental animals was 5%, based on decreased growth and increased cecal weights at 15% beta-glucan, and the NOEL for embryotoxicity was also 5%, based on decreased weight gain during lactation in the 15% beta-glucan group.

The teratogenic potential of beta-glucan (as curdlan) was determined using groups of 15-20 gravid Dutch belted rabbits.[42] The rabbits were dosed orally with 0, 1, 2, or 5 g/kg bw/day beta-glucan in a gelatin capsule delivered using a syringe. The 5 g/kg dose was administered as two divided doses, and the controls received two empty capsules. The rabbits were killed on day 28 of gestation. None of the controls died, but one, one, and three dams of the 1, 2, and 5 g/kg bw/day groups, respectively, died during the study. Eleven resorptions were observed in the high dose group, as compared to four in the control group, six in the 1 g/kg group, and five in the 2 g/kg group. The researchers stated, however, that the number of dams with resorptions was similar in all groups, and that no teratogenic effects were observed. The NOEL for both maternal and embryotoxicity was 5 g/kg bw/day.

GENOTOXICITY

Genotoxicity studies are summarized in Table 10. The in vitro genotoxicity of gellan gum (≤20 mg/ml), sodium dextran sulfate (≤25 mg/plate), beta-glucan (≤5000 μg/plate or /ml), sodium carboxymethyl beta-glucan (≤50,000 μg/plate or /ml) and pullulan (≤12 mg/ml) was evaluated using Ames test, chromosomal aberration assays, and/or DNA repair tests, with and without metabolic activation. The results were negative in these tests. The only non-negative result was a weak positive outcome with 20 mg/plate pullulan in a rec assay using *Bacillus subtilis*. Negative results were also reported in in vivo mouse micronucleus tests with ≤5000 mg/kg beta-glucan and ≤1800 mg/kg pullulan.

CARCINOGENICITY

Oral

Gellan Gum

Groups 50 male and 50 female Swiss Crl mice were fed a diet containing 0, 1, 2, or 3% gellan gum (varied degree of acetylation; 58.5% polysaccharide) for 96 wks (males) or 98 wks (females).[39] No treatment-related effects on body weights or feed consumption were observed. No treatment-related neoplastic or non-neoplastic lesions were reported.

In another study, groups of 50 male and 50 female F_1 generation Sprague-Dawley rats were exposed *in utero* to gellan gum, and were maintained on a diet containing 0, 2.5, 3.8, or 5.0% gellan gum (varied degree of acetylation; 58.5% polysaccharide) for 104 wks.[39] Survival of treated male rats was decreased when compared to controls, but survival of treated female rats was better than the concurrent controls. Male rats of the 3.8 and 5.0% test groups had decreased body weights compared to controls initially and after 76 wks. However, the researchers stated that the growth pattern of these test animals was the same as that of the controls, and the lower body weights were not indicative of toxicity. There were no neoplastic or non-neoplastic lesions associated with dosing, and gellan gum was not carcinogenic when fed to Sprague-Dawley rats.

Dextran

A group of 15 male and 15 female ACI rats were fed a diet containing 2.5% dextran (21,500 MW) for 480 days, and the control group of 20 males and 20 females was fed a basal diet.[62] Body weights gains of treated male rats were statistically significantly decreased compared to controls. An increased incidence in tumors was not reported, and no intestinal tumors were found.

Sodium Dextran Sulfate

A number of studies have demonstrated that oral exposure to sodium dextran sulfate produces colon cancer in rats; the mechanism is not genotoxic. Oral administration has been shown to induce colonic inflammation, and a 2-day study in which female Fischer 344 rats were given 3 or 6% sodium dextran sulfate in the drinking water indicated that oxidative DNA damage occurred in the colonic mucosa.[63] The MW of sodium dextran sulfate has been found to be a factor in carcinogenic activity. While an extensive number of studies are available in the published literature, just a few are summarized below. An example of an inflammation-related mouse colon carcinogenesis model is described, with an indication of strain-dependency.

In one study evaluating the carcinogenic potential of sodium dextran sulfate (54,000 avg. MW; sulfur content 18.9%) in ACI rats, 10 males were fed a diet containing 10% sodium dextran sulfate, 14 males and 12 females were fed 5% in the diet, and a control group of nine males and nine females were given a basal diet.[64] All animals were necropsied at natural death or when killed due to moribund condition. All animals in the 10% group died 6-14 days after initiation of dosing, and all had severe acute nephrosis. Two animals of the 5% group died on day 14, but most of the remainder of this group lived for more than 130 days. Blood was observed on the surface of the stools of these animals at 2.5 mos. The weight gain of animals of this group was decreased compared to controls. Of the 23 rats that survived for more than 130 days, 15 rats developed intestinal tumors; tumors included five adenomas, five adenocarcinomas, and three papillomas in the colon and six adenomas and two adenocarcinomas in the cecum. While most rats had a single tumor upon gross observation, microscopic examination found multicentric foci of atypical hyperplasia of the glandular epithelium. No intestinal tumors were reported in the control group.

In a second study testing the same sodium dextran sulfate, 15 male and 15 female ACI rats were fed 1% in the diet for 660 days; the avg. daily intake was 0.15 g/day/animal.[65] A control group of 10 males and 10 females were fed a basal diet. All but two treated male rats survived for 350+ days. Body weight gains of the test group were similar to that of the controls. Intestinal tumors were observed in 22 treated rats; 16 papillomas, four squamous cell carcinomas, two adenomas, and five adenocarcinomas were reported in the colon and rectum and one adenocarcinoma was found in the cecum. Thirteen tumors were reported at miscellaneous sites. Again, no intestinal tumors were reported in the control group.

To examine the effect of MW, three groups of 15 male and 15-16 female ACI rats were fed for 480 days a diet containing 2.5% of a sodium dextran sulfate, MW 520,000, 54,000, or 9500; each sodium dextran sulfate had a sulfur content of 18-19%.[62] (The 54,000 MW substance was synthesized using the dextran described previously.) A control group of 20 male and 20 female rats was fed a basal diet. There was no significant difference in survival time between any of the groups. Body weights gains of male rats of the 54,000 MW diet were statistically significantly decreased compared to control males; body weight gains in the other two groups were similar to controls. The 54,000 MW sodium dextran sulfate had the strongest carcinogenic activity, with tumors

being reported similar to the studies described previously. The other two MW substances did not show significant carcinogenic activity; only 2 colorectal adenocarcinomas were observed in the 520,000 MW group. Colorectal squamous metaplasia was observed in most rats in all test groups. Other miscellaneous tumors were reported, but there was no statistically significant difference between treated and control groups.

A colitis-related mouse colon carcinogenesis model was developed.[66] When eight male Crj:CD-1 mice were given a single i.p. injection of 10 mg/kg azoxymethane (AOM), followed by 7 days of 2% sodium dextran sulfate in the drinking water 1 wk later, there was a 100% incidence of colonic adenocarcinomas and 38% of adenomas at wk 20. No adenocarcinomas were observed in any of the mice dosed with AOM and sodium dextran sulfate simultaneously, 1 wk of sodium dextran sulfate and then AOM, AOM only, or sodium dextran sulfate only, and only two adenomas (in 10 mice) were observed when AOM was given during sodium dextran sulfate administration. All of the adenocarcinomas were positive for β-catenin, cyclooxygenase (COX)-2, and inducible nitric oxide synthase (iNOS) and negative for immunoreactivity of p53.

Strain-sensitivity to the above model was then examined.[67] In testing with male Balb/c, C3H/HeN, C57BL/6N, and DBA/2N mice, Balb/c mice were very sensitive to the model. C57BL/6N mice also developed colonic adenocarcinomas, but to a lesser extent. C3H/HeN and DBA/2N mice developed only a few colonic adenomas. However, the greatest inflammation response was observed in C3H/HeN mice, followed by Balb/c mice. The researchers hypothesized that the strain differences in susceptibility of colon carcinogenesis induced by AOM and sodium dextran sulfate might be influenced by the response to nitrosation stress due to inflammation as determined by the genetic background.

Anti-Tumor Effects

Xanthan Gum

Groups of C57BL/6 mice were inoculated subcutaneously (s.c.) with 1 x 10^6 B16K[b] melanoma cells.[68] A suspension of 10 mg/ml xanthan gum, 100 µl, or PBS was given by gavage once every 5 days, starting 1 day prior to inoculation. Rapid tumor growth occurred in PBS-treated mice, but tumor growth was statistically significantly reduced in xanthan gum-treated mice. Spleen cell composition was analyzed 22 days after inoculation. There was no change in overall cellular composition, but natural killer (NK) cells and tumor-specific cytotoxic T-lymphocyte (CTL) activity were increased by xanthan gum. All PBS-treated mice died by day 46 after inoculation; 40% of the xanthan gum-treated mice were alive at day 100. The researchers demonstrated that the anti-tumor effect of xanthan gum was highly dependent on Toll-like receptor (TLR) 4-mediated signaling.

Pullulan

Male Balb/c mice were used to determine the anti-tumor and anti-metastatic potential of pullulan (water-soluble low-MW β-(1→3)- with 50-80% branched β-(1→6); MW 100,000).[69] Colon-26 cells were implanted into the spleens of mice on day 0, and groups of mice were dosed orally with 25 or 50 mg/kg or i.p. with 5 or 15 mg/kg pullulan for 14 consecutive days, starting 12 h after implantation. Control groups consisted of sham-operated mice (not implanted with colon-26 cells) and mice implanted with the cells but given physiological saline or distilled water instead of pullulan. Splenic tumor weights were reduced with the 50 mg/kg oral and 15 mg/kg i.p. doses of pullulan, but not with the other doses. Liver metastasis was significantly inhibited with 50 (but not 25) mg/kg oral and 5 and 15 mg/kg i.p. pullulan.

Levan

The anti-tumor activity of levan produced from four different microorganisms, i.e., *Gluconoacetobacter xylinus*, *Rahnella aquatilis*, *Zymomonas mobilis*, and *Microbacterium laevaniformans* (G-levan, R-levan, Z-levan, and M-levan, respectively) was evaluated using female ICR mice.[70] The MWs of these levans were 40,000, 380,000, 570,000, and 710,000, respectively. On day 0, 0.2 ml (equiv. to 3 x 10^6 cells) of sarcoma-180 tumor cells were implanted s.c. into the groin of the mice. On days 1 through 7, 200 mg/kg of the levans in distilled water were applied to the mice, and the mice were killed on day 26. The tumor growth inhibition ratio (IR) ranged from 42.2-69.6% for the four levans. M-Levan had the highest I.R. value, but it was not statistically significant. The IRs for R- and Z-levan were comparable, and the G-levan was significantly less effective.

IRRITATION AND SENSITIZATION

Non-human and human dermal irritation and sensitization studies are summarized in Table 11. The dermal irritation and sensitization potentials of xanthan gum, beta-glucan, and sodium carboxymethyl beta-glucan were evaluated in animal studies. Xanthan gum, up to 1%, and beta-glucan, concentration not specified, were not irritating to rabbit skin. One study with 5% aq. xanthan gum on shaved rabbit skin produced localized irritation; study details were not provided. Neither xanthan gum, tested at 0.1%, nor beta-glucan (concentration not specified) were sensitizers in guinea pigs. Sodium carboxymethyl beta-glucan was at most a slight skin irritant in guinea pigs at a concentration of 50% aq.; a 10% aq. solution was not a sensitizer in guinea pigs.

In humans, neither beta-glucan nor sodium carboxymethyl beta-glucan were irritants or sensitizers. The test concentration of beta-glucan was not specified. Sodium carboxymethyl beta-glucan was applied neat in the irritation study and as a 2% aq. solution in the sensitization study.

Phototoxicity

A human photoallergenicity study also is summarized in Table 11. A 2% aq. solution of sodium carboxymethyl beta-glucan was not photosensitizing in clinical studies.

Adverse Reactions

Dextran

Over a 5-yr period (1981-1986), 12,646 dextran 70 units were administered to 5745 patients (mean of 2.2 units/patient) undergoing gynecological surgery or a Cesarean section as a plasma volume expander.[71] Fifteen immediate dextran-induced anaphylactoid reactions were reported, with an incidence of one reaction/383 patients treated. Life-threatening reactions occurred in 7 of these patients.

Ocular Irritation

Ocular irritation studies are summarized in Table 12. Xanthan gum, 1%, and up to 0.8% gellan gum were not ocular irritants in rabbit eyes, and up to 0.5% gellan gum was not irritating to human eyes. The ocular irritation potential of sodium carboxymethyl beta-glucan was described as weakly irritating in a HET-CAM assay, but was practically non-irritating in rabbit eyes.

SUMMARY

Microbial polysaccharide gums can be produced intercellularly, by the cell wall, or exocellularly. Exocellular polysaccharide gums constantly diffuse into the cell culture medium and are easily isolated, while cell wall and intercellular polysaccharide gums are more difficult to separate from cell biomass. Many of the 34 microbial polysaccharide gums discussed in this safety assessment are produced exocellularly. They are reported to have numerous functions in cosmetics, including emulsion stabilizer, film former, binder, viscosity increasing agent, and skin conditioning agent.

The same microbial polysaccharide gums can often be produced by more than one organism. For example, beta-glucan can be produced by fungi, yeasts, and grains and levan can be produced by bacteria, yeasts, or fungi. The properties of the microbial polysaccharide gums can vary widely based on, among other parameters, the side groups, the ester substituents, or bacterial strains. These polysaccharide gums are generally very large molecules, and the MW of each ingredient can vary considerably.

Xanthan gum is reported to be used in almost every category of cosmetic product, with 3470 reported uses. Biosaccharide gum-1, sclerotium gum, and beta-glucan are reported to be used in 346, 193, and 137 cosmetic formulations, respectively. All other in-use ingredients have less than 70 uses. The ingredient with the highest concentration of use is pullulan; it is used at up to 12% in leave-on formulations and 17% in a breath freshener. Both xanthan gum and biosaccharide gum-1 are used at up to 6% in leave-on formulations and xanthan gum crosspolymer and biosaccharide gum-4 are used at 5% in leave-on formulations. All other in-use ingredients are used at concentrations of ≤3%.

Xanthan gum, gellan gum, and beta-glucan are approved as direct food additives, and xanthan gum and dextran are approved indirect food additives. Xanthan gum and dextran also have pharmaceutical applications.

Xanthan gum, orally administered, is slowly broken down in the gut by enzymatic and non-enzymatic mechanisms, and can be absorbed in some form to some extent. The absorbed fraction does not accumulate in the tissues and can be completely metabolized to CO_2. Gellan gum, orally administered, does not breakdown to any substantial extent in the gut, and is only very poorly absorbed.

Dextrans, 3,000-20,000 MW, are not absorbed through intact skin in humans. They are absorbed through the dermis (up to 38% for 3000 MW) if the epidermis is removed (e.g., via mini-erosion); absorption in this case is inversely proportional to MW. Orally administered dextrans, 4400 to 40,500 MW, are not absorbed to any appreciable extent in the gut (very low bioavailability). If dextrans were absorbed to a significant extent through the skin, animal studies in which dextrans were administered i.v. indicate that their half-lives in blood plasma would be directly related to the MW; they would be excreted in the urine to an extent that is inversely related to the MW; the lower MW dextrans would tend to be readily excreted unchanged in the urine; and the higher MW dextrans would have the potential to accumulate and to be broken down in the liver and other tissues, and would be more likely to be excreted in the urine in a dose-dependent manner than the lower MW dextrans. Dextran sulfate, 7000 to 8000 MW, orally administered, is only very poorly absorbed in the human GI tract. After i.v. administration, it is rapidly excreted, mostly intact, in the urine, although at least some of it can accumulate in the tissues (to a substantially greater extent than observed after oral exposure), and some of it appears to be incorporated into glycogen and other substances in the body.

Beta-glucan in a topically applied solution can penetrate into the epidermis and dermis. There appears to be no information about its absorption through the skin into the bloodstream. Orally administered, it is readily metabolized to CO_2, at least partially by microflora in the gut. Beta-glucan is not well absorbed or eliminated after i.p. injection. Pullulan, orally administered, is hydrolyzed to some small extent in the gut. It is partially hydrolyzed by amylases in the upper GI tract of human subjects, and is subject to breakdown by intestinal microflora to form short-chain fatty acids; the rate of the latter depends on the degree of polymeriza-

tion. However, pullulan can be essentially completely broken down to short-chain fatty acids in the human gut. It has not been determined to what extent, if any, the products of hydrolysis can be absorbed.

The acute toxicity of xanthan gum, gellan gum, beta-glucan, sodium carboxymethyl beta-glucan, and pullulan was assessed orally in mice, rats, and/or dogs, and dextran sulfate and beta-glucan were tested by i.p. and i.v. dosing in mice and rats. There was no notable toxicity observed in these studies. In acute inhalation studies, the LC_{50} of xanthan gum was >21 mg/l in rabbits and of gellan gum was >5.06 mg/l in rats. A single 180-min exposure of humans office dust contaiing 10 mg curdlin/g dust resulted in decreased nasal volume, in swelling in the nasal turbinates, and an increase in nasal eosinophils when compared to "clean" dust.

The inflammatory response following a single exposure to beta-glucan (as curdlan) and to pullulan was also evaluated in guinea pigs. Mostly, no effect or a slight decrease in inflammatory cells in lung lavage was observed. A 4 h inhalation exposure to beta-glucan in dust by guinea pigs produced a delayed subacute nasal congestion when compared to dust without beta-glucan and resulted in decreased nasal volume. Industrial exposure to xanthan gum powder did not appear to cause significant acute or chronic pulmonary effects. Using artificial skin, 5% levan had an anti-inflammatory effect in irritated skin.

Repeated dose oral toxicity of xanthan gum was evaluated in rats and dogs, of gellan gum in rats, dogs, and monkeys, of dextran in rats, of beta-glucan in mice, rats, and dogs, and of pullulan in rats. Most of the studies were dietary, and study durations lasted up to 2 yrs. Most observations were related to changes in feed consumption and intestinal effects. Inhalation exposure to 100 µg/ml beta-glucan (as curdlan), 4 h/day, 5 days/wk for 4 wks, did not have an effect on the cells of the lung lavage or cell wall, and there were no microscopic lesions of the lung. With i.p. administration, no toxicity was reported when mice were dose 10 times over 2 wks with 5 mg xanthan gum in 0.5 ml water or mice or guinea pigs were dosed with 250 mg/kg bw beta-glucan for 7 days. Intravenous administration of 40 and 1000 mg/kg bw beta-glucan for 30 days resulted in hepatosplenomegaly in mice.

No toxic effects were observed in human subjects with oral ingestion of 150 mg/kg/day xanthan gum or 175-200 mg/kg/day gellan gum for 23 days, 10 g of dextran or pullulan for 14 days, or 6-50 g/day beta-glucan for up to 28 days. No significant or chronic effects in pulmonary function were reported in groups exposed occupationally to xanthan gum.

Dietary reproductive and developmental toxicity studies were conducted with xanthan gum, gellan gum, and beta-glucan. No reproductive or developmental effects were reported in a three-generation reproductive study in which rats were fed diets containing up to 5 g/kg bw/day xanthan gum. Gellan gum, up to 5%, did not have a fetotoxic or teratogenic effect on rats. Dietary administration of beta-glucan in rats in reproductive and developmental studies did not have any reproductive effects, but there were statistically significant decreases in body weights and body weight gains in offspring and parental animals. In a teratogenicity study in which rabbits were dosed with 5 g/kg bw/day beta-glucan, an increase in resorptions in the 5 g/kg group was considered similar to the other test groups and the controls, and 5 g/kg beta-glucan was not teratogenic in rabbits.

The in vitro genotoxicity of gellan gum (≤20 mg/ml), sodium dextran sulfate (≤25 mg/plate), beta-glucan (≤5000 µg/plate or /ml), sodium carboxymethyl beta-glucan (≤50,000 µg/plate or /ml) and pullulan (≤12 mg/ml) was evaluated in Ames test, chromosomal aberration assays, and/or DNA repair tests, with and without metabolic activation. Results were negative in all of these tests. The only non-negative result was a weak positive result with 20 mg/plate pullulan in a rec assay using *Bacillus subtilis*. Negative results were also reported in in vivo mouse micronucleus tests with ≤5000 mg/kg beta-glucan and ≤1800 mg/kg pullulan.

Dietary studies examining the carcinogenic potential of ≤5% gellan gum and 2.5% dextran reported that neither of these ingredients caused an increase in tumors. However, a number of studies have demonstrated that oral exposure to sodium dextran sulfate produces colon carcinogenesis in rats, the mechanism is non-genotoxic. In one study, the MW of sodium dextran sulfate was a factor in carcinogenic activity; a 54,000 MW sodium dextran sulfate produced colorectal tumors, but 9500 and 520,000 MW sodium dextran sulfate did not have significant carcinogenic activity. Oral administration has been shown to induce colonic inflammation, and a 2-day study in which female Fischer 344 rats were given 3 or 6% sodium dextran sulfate in the drinking water indicated that oxidative DNA damage occurred in the colonic mucosa. An inflammation-related mouse colon carcinogenesis model indicated that the development of colonic tumors is strain-dependent, and that Balb/c mice were very sensitive to the model. C57BL/6N mice also developed tumors, but to a lesser extent, while C3H/HeN and DBA/2N mice only developed a few tumors.

Oral administration of 10 mg/ml xanthan gum had an anti-tumor effect in mice inoculated with melanoma cells, and 50 mg/kg pullulan, but not 15 mg/kg, significantly inhibited tumor growth in mice following implantation of colon-26 cells. Levan did not have an anti-tumor effect in mice.

Non-human and human dermal irritation and sensitization studies are summarized in Table 11. The dermal irritation and sensitization potentials of xanthan gum, beta-glucan, and sodium carboxymethyl beta-glucan were evaluated in animal studies. Xanthan gum, up to 1%, and beta-glucan, concentration not specified, were not irritating to rabbit skin. One study with 5% aq. xanthan gum on shaved rabbit skin produced localized irritation; however, study details were not provided. Neither xanthan gum, tested at 0.1%, nor beta-glucan (concentration not specified) were sensitizers in guinea pigs. Sodium carboxymethyl beta-glucan was at most a slight skin irritant in guinea pigs at a concentration of 50% aq.; a 10% aq. solution was not a sensitizer in guinea pigs.

In humans, neither beta-glucan nor sodium carboxymethyl beta-glucan were irritants or sensitizers. The test concentration of beta-glucan was not specified. Sodium carboxymethyl beta-glucan was applied neat in the irritation study and as a 2% aq. solution in the sensitization study. A 2% aq. solution of sodium carboxymethyl beta-glucan was not photosensitizing in clinical studies.

Xanthan gum, 1%, and gellan gum, up to 0.8%, were not ocular irritants in rabbit eyes, and up to 0.5% gellan gum was not irritating to human eyes. The ocular irritation potential of sodium carboxymethyl beta-glucan was described as weakly irritating in a HET-CAM assay, but the ingredient was practically non-irritating in rabbit eyes.

DISCUSSION

Microbial polysaccharide gums are produced by a wide variety of microorganisms, and some can also be isolated from plants, e.g., beta-glucan can be isolated from barley and oats. Although these ingredients are produced primarily by microbial sources, the cosmetic ingredients are purified during manufacture, thus microbial contamination is not a concern.

The Panel determined that, even though there are some data gaps, the available data on polysaccharide gums included in this safety assessment may be extrapolated to support the safety of the entire group. And while there was no specific data on the hydroxy-propyltrimonium chloride compounds, data included in a previous CIR safety assessment on trimonium ingredients are applicable in determining the safety of the three hydroxypropyltrimonium chloride compounds included in this report.

Parenterally administered polysaccharides appear to be biotransformed to a limited but variable extent in animal and human studies. However, these very large compounds appear not to be significantly absorbed through the skin and would have negligible bioavailability. Coupled with a lack of significant toxicity associated with other routes of exposure, the CIR Expert Panel determined that systemic effects were unlikely to result from topical application of cosmetics containing these ingredients.

The Panel initially expressed concern for a study that reported 5% aq. xanthan gum caused irritation. However, this was the only finding of irritation among almost 20 studies on microbial polysaccharide gums. Given the absence of study details, including no mention of a control group, it appeared to the Panel to be more likely that the irritation was the result of the study methodology (e.g., shaved skin) and not by the xanthan gum. There was no evidence of sensitization in human or non-human testing.

The Panel also remarked on the induction of colon cancer in rodents with oral exposure to sodium dextran sulfate. Sodium dextran sulfate is a commonly used model for induction of colitis in a well-characterized mouse model to study colitis, and while the mode of action is unknown it is not relevant to human cosmetic exposure.

Finally, the Panel discussed the issue of incidental inhalation exposure to microbial polysaccharide gums from powders and other products that may be aerosolized. These ingredients are reportedly used at concentrations up to 1% in cosmetic products that may be aerosolized and up to 6% in other products that may become airborne. The Panel noted that 95% – 99% of droplets/particles would not be respirable to any appreciable amount. Furthermore, droplets/particles deposited in the nasopharyngeal or bronchial regions of the respiratory tract present no toxicological concerns based on the chemical and biological properties of this ingredient. Coupled with the small actual exposure in the breathing zone and the concentrations at which the ingredients are used, the available information indicates that incidental inhalation would not be a significant route of exposure that might lead to local respiratory or systemic effects. Results of acute inhalation studies with xanthan gum and gellan gum produced no significant toxicity, nor did results from a 4-wk inhalation study of beta-glucan in guinea pigs. Industrial exposure to xanthan gum powder caused no significant acute or chronic effects in pulmonary function. The Panel also considered the data available to characterize the potential for microbial polysaccharide gums to cause systemic toxicity, irritation, sensitization or other effects. The Panel noted that testing with a number of microbial polysaccharide gums demonstrated they did not produce systemic toxicity in oral studies; they are not reproductive or developmental toxicants; are not genotoxic; and are not considered irritants or sensitizers. A detailed discussion and summary of the Panel's approach to evaluating incidental inhalation exposures to ingredients in cosmetic products is available at http://www.cir-safety.org/cir-findings.

CONCLUSION

The CIR Expert Panel concluded the microbial polysaccharide gums listed below are safe in the present practices of use and concentration in cosmetics.

Xanthan Gum	Biosaccharide Gum-3*
Hydroxypropyl Xanthan Gum*	Biosaccharide Gum-4
Undecylenoyl Xanthan Gum*	Biosaccharide Gum-5*
Dehydroxanthan Gum	Pseudoalteromonas Exopolysaccharides*
Xanthan Gum Crosspolymer	Dextran
Xanthan Hydroxypropyltrimonium Chloride*	Carboxymethyl Dextran*
Gellan Gum	Dextran Hydroxypropyltrimonium Chloride*
Welan Gum*	Sodium Carboxymethyl Dextran
Biosaccharide Gum-1	Dextran Sulfate
Biosaccharide Gum-2	Sodium Dextran Sulfate

Sclerotium Gum   Sodium Carboxymethyl Beta-Glucan
Hydrolyzed Sclerotium Gum   Pullulan
Beta-Glucan   Myristoyl Pullulan*
Beta-Glucan Hydroxypropyltrimonium Chloride*   Levan*
Beta-Glucan Palmitate*   Rhizobian Gum
Hydrolyzed Beta-Glucan*   Hydrolyzed Rhizobian Gum
Oxidized Beta-Glucan*   Alcaligenes Polysaccharides

Were ingredients in this group not in current use (as indicated by *) to be used in the future, the expectation is that they would be used in product categories and at concentrations comparable to others in the group.
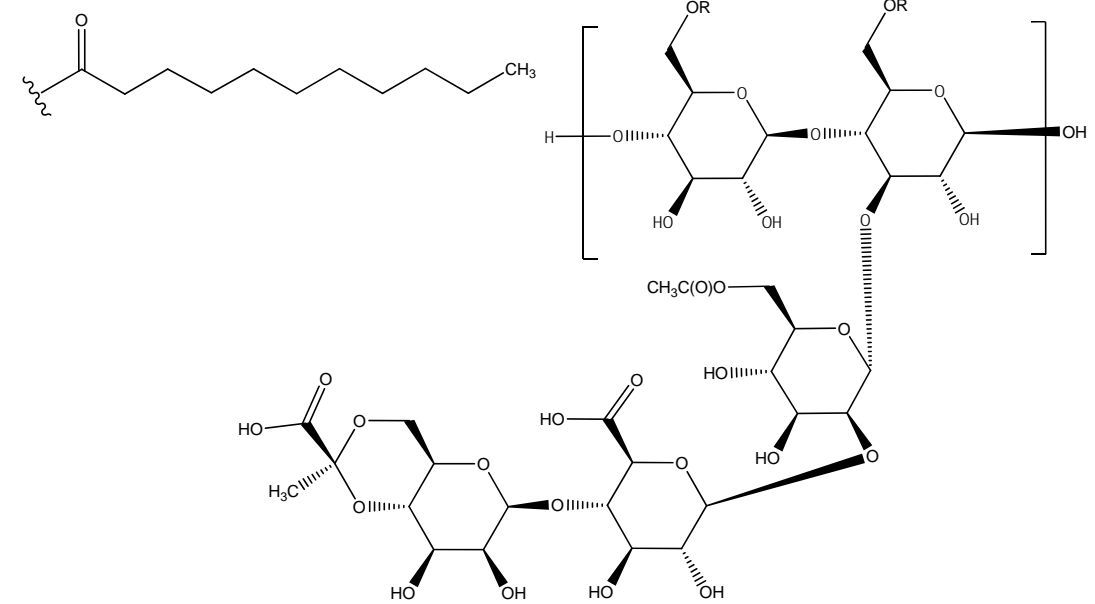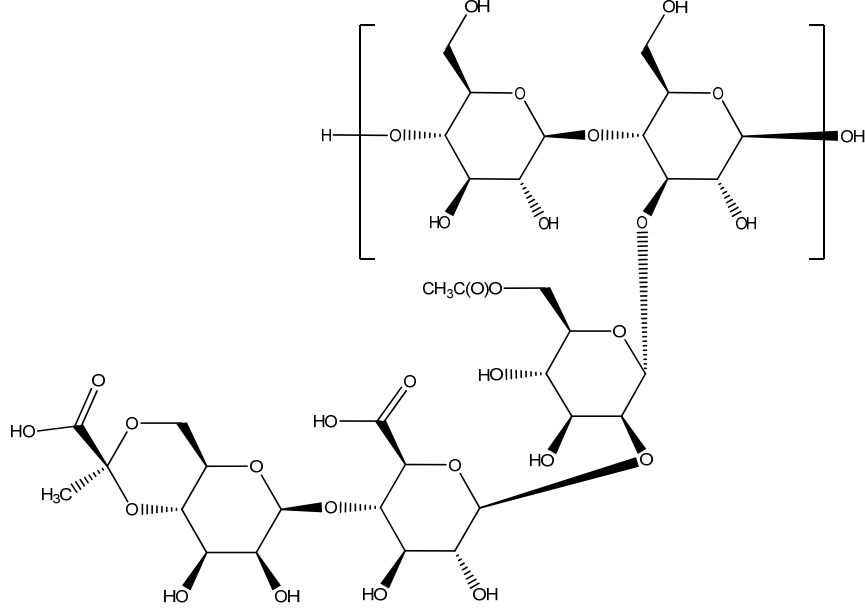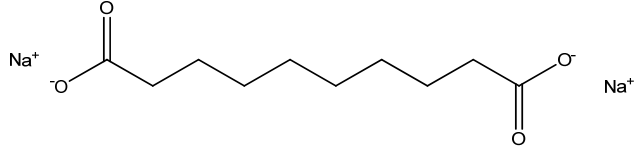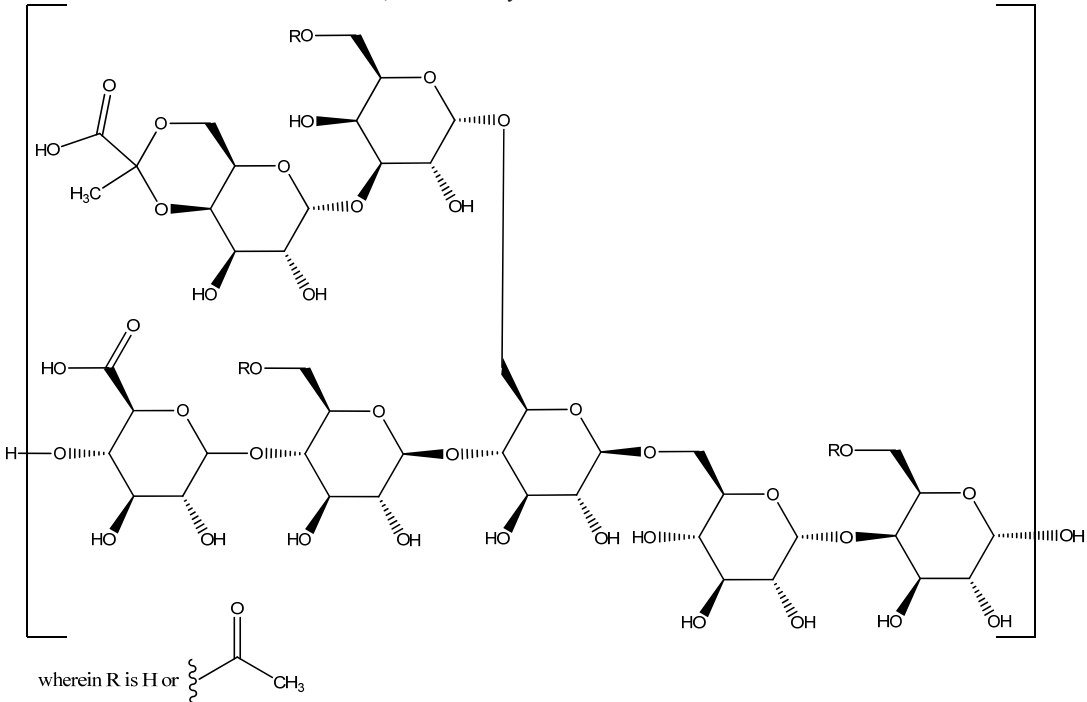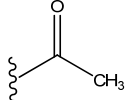
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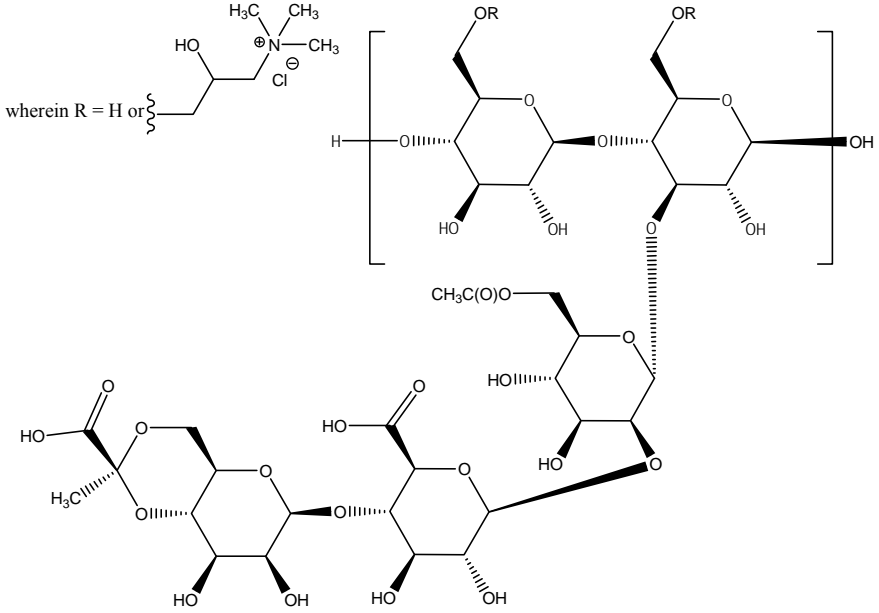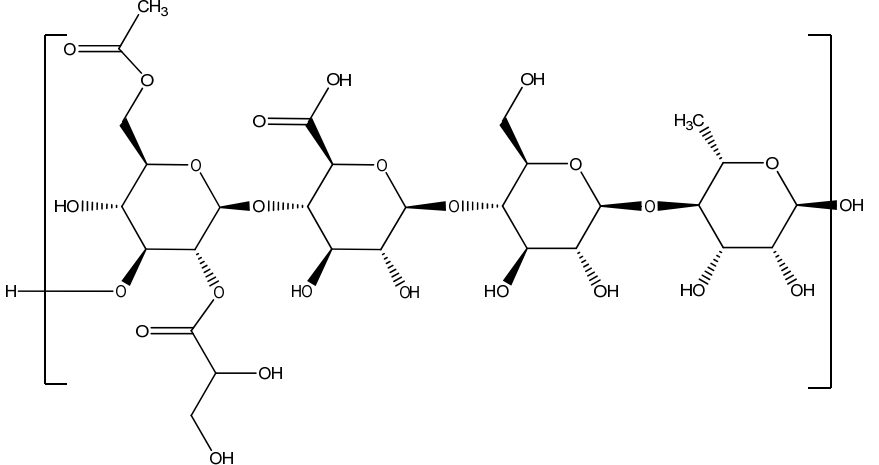
Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Xanthan Gum 11138-66-2	a high MW heteropolysaccharide gum produced by a pure-culture fermentation of a carbohydrate with *Xanthomonas campestris*;[14] composed of glucose, glucuronic acid, 6-acetylmannose, and 4,6-pyruvylated mannose residues[72]	binder; emulsion stabilizer; skin conditioning agent-misc.; surfactant-emulsifying agent; viscosity increasing agent-aq.	(given below definition)

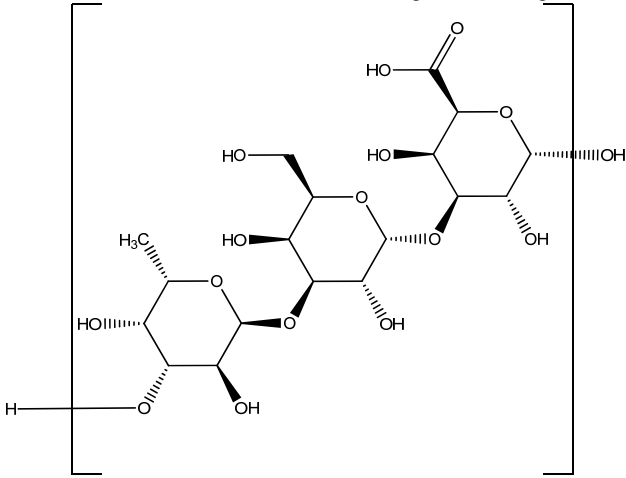| Hydroxypropyl Xanthan Gum 106442-37-9 | the hydroxypropyl ether of xanthan gum,[14] wherein ether substitution occurs primarily at 6-position of the backbone sugar residues | emulsion stabilizer; film former; viscosity increasing agent-aq. | |
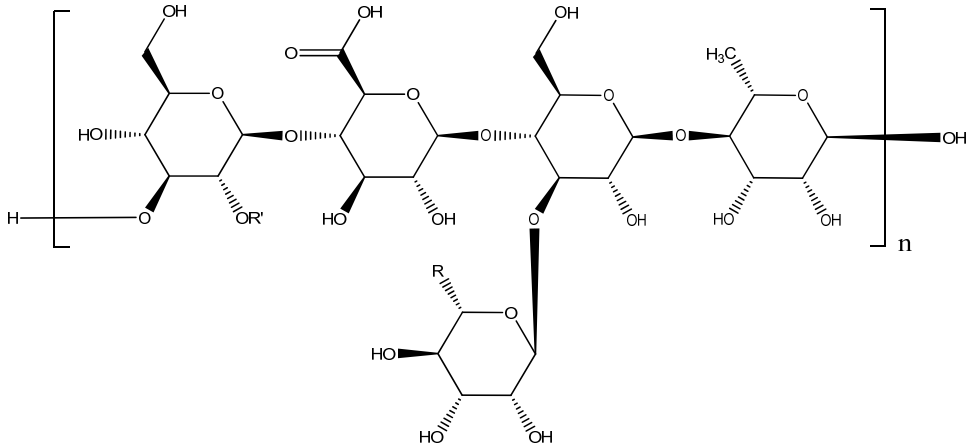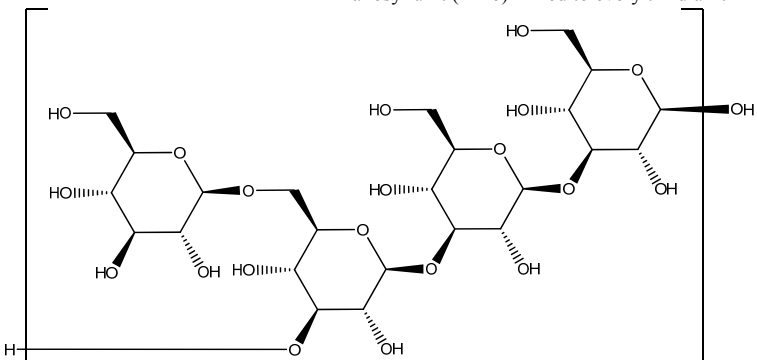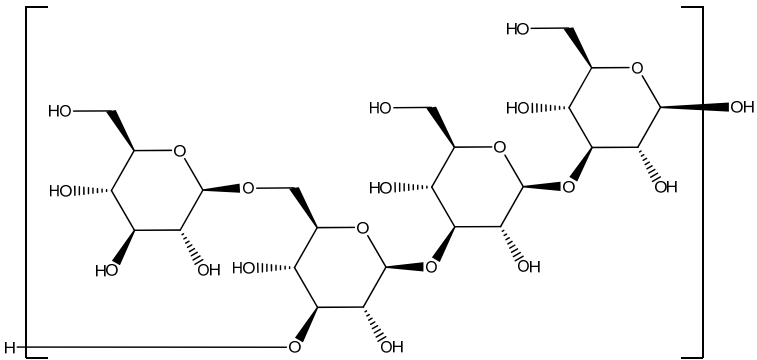
wherein R = H or

16

Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Undecylenoyl Xanthan Gum	the condensation product of undecylenic acid chloride and xanthan gum,[14] wherein esterification occurs primarily at the 6-position of backbone sugar residues	emulsion stabilizer; hair conditioning agent	

wherein R = H or

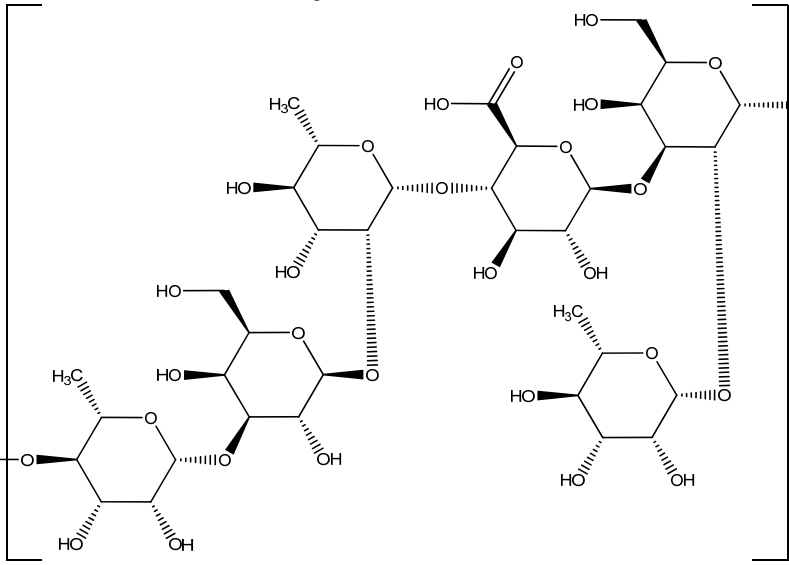| Dehydroxanthan Gum 11138-66-2 [dehydro is co-listed under this CAS No. with Xanthan Gum] | the product obtained by the dehydration of xanthan gum[14] | emulsion stabilizer; film former; hair fixative; suspending agent-nonsurfactant; viscosity increasing agent-aq. | |
| Xanthan Gum Crosspolymer | xanthan gum crosslinked with disodium sebacate[14] | skin protectant; skin conditioning agent-misc. | |

Crosslinked with:

Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Xanthan Hydroxypropyl Trimonium Chloride	the quaternary ammonium salt formed by the reaction of Xanthan Gum with 2,3-epoxypropyltrimethylammonium chloride	hair conditioning agent; skin conditioning agent-humectant; viscosity increasing agent-aq	

wherein R = H or

Other Gums			
Gellan Gum 71010-52-1	a high MW heteropolysaccharide gum produced by pure-culture fermentation of a carbohydrate with *Pseudomonas elodea*;[14] consists of the basic backbone →3)-β-D-Glc-(1→4)-β-D-GlcA-(1→4)-β-D-Glc-(1→4)-α-L-Rha-(1→);[11] composed of glucose (some of which is glyceryl and/or acetyl esterified), glucuronic acid, and rhamnose residues[12]	emulsion stabilizer; viscosity increasing agent-aq.	

Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Welan Gum 96949-22-3	a gum produced by the fermentation of *Alcaligenes*;[14] consists of the basic backbone →3)-β-D-Glcp-(1→4)-β-D-GlcpA-(1→4)-β-D-Glcp-(1→4)-α-L-Rhap-(1→3)↖1-α-L-Rhap **or** α-L-Manp; composed of glucose, glucuronic acid, rhamnose and mannose residues, with rhamnopyranosyl and/or L-mannopyranosyl side chains[73]	binder; emulsion stabilizer; film former; viscosity increasing agent-aq.	

wherein R is CH_3 or CH_2OH and R' is H or $C(O)CH_3$

Sclerotium Gum 39464-87-4	the polysaccharide gum produced by the fungus, *Sclerotium rolfsii*;[14] consists of the basic backbone of β-D-(1→3)-glucopyranosyl units with a β-D-glucopyranosyl unit (1→6) linked to every third unit[74]	emulsion stabilizer; skin conditioning agent-misc.; viscosity increasing agent-aq.	

Hydrolyzed Sclerotium Gum	the partial hydrolysate of sclerotium gum derived by acid, enzyme, or other method of hydrolysis[14]	film former; skin protectant; skin conditioning agent-humectant	

Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Rhizobian Gum 127712-52-1 [for Rhizob*ium* Gum]	the polysaccharide gum produced by the fermentation by *Rhizobium bacterium*;[14] composed of glucose, glucuronic acid, galactose and pyruvylated galactose residues, with some acylation[75]	film former; hair fixative; plasticizer; suspending agent-nonsurfactant; viscosity increasing agent-aq	

wherein R is H or

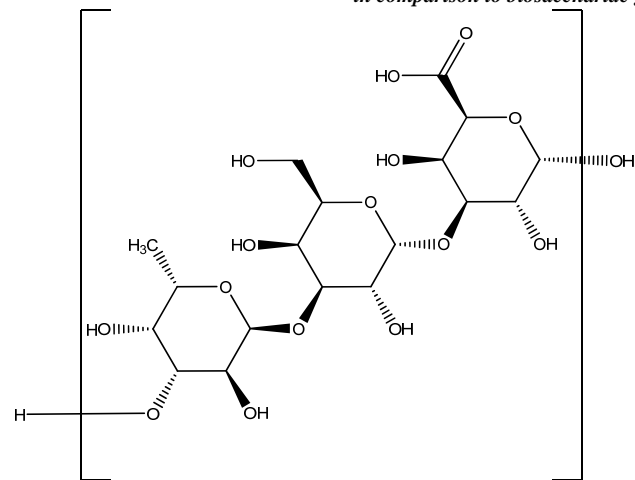| Hydrolyzed Rhizobian Gum	the partial hydrolysate of Rhizobian Gum derived by acid, enzyme, or other method of hydrolysis[14]	film former	
Biosaccharide Gum-1 223266-93-1	a fermentation gum derived from sorbitol; described as a polymer of α-L-fucose(1→3)α-D-galactose(1→3)-α-D-galacturonic acid trisaccharides; [14] composed of fucose, galactose, and galacturonic acid residues	skin conditioning agent-misc.	

Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Biosaccharide Gum-2 758716-52-8	a fermentation gum derived from sorbitol; described as a polymer of α-L-Rhap(1→3)-β-D-Galp-(1→2)-α-L-Rhap-(1→4)-β-D-GlepA (1→3)-[α-L-Rhap-(1→2)-]-α-D-Galp-(1;[14] composed of rhamnose, galactose, and glucuronic acid residues	skin conditioning agent-misc.	

Biosaccharide Gum-3 896736-76-8	a fermentation gum derived from sorbitol; described as a polymer of α-L-fucose(1"3)-α-D-galactose(1"3)-α-D-galacturonic acid trisaccharides, ***and is characterized by a smaller degree of polymerization and MW in comparison to biosaccharide gum-1***[14]	skin conditioning agent-misc.	
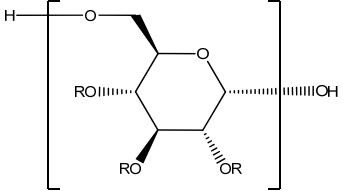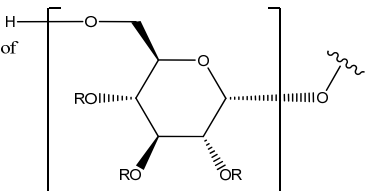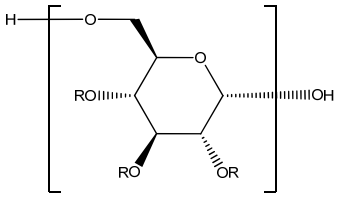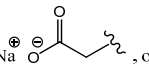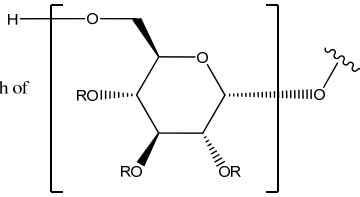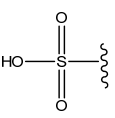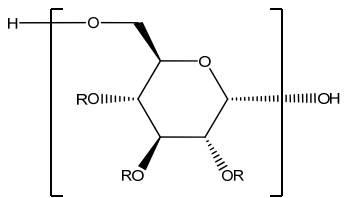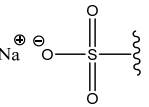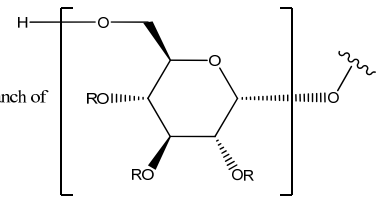
Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Biosaccharide Gum-4 283602-75-5	a fermentation gum derived from sorbitol; it is a de-acetylated branched polymer consisting of L-fucose, 2-D-glucose and glucuronic acid repetitive units[14]	skin conditioning agent-misc.	
Biosaccharide Gum-5	a fermentation gum derived from sorbitol; described as a polymer of –glucose-β-(1-->2)-rhamnose-α-(1-->4)-glucose-α-(1-->3)-rhamnose-β-(1-->4), with rhamnose –α-(1-->3) branching[14]	skin conditioning agent-misc.	
Pseudoalteromonas Exopolysaccharides	the exopolysaccharide excreted by the fermentation of *Pseudoalteromonas*	skin conditioning agent-misc.	

wherein R is H or

Table 1. Definition, Function, and Idealized Structure
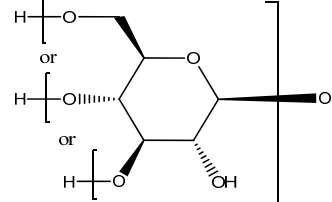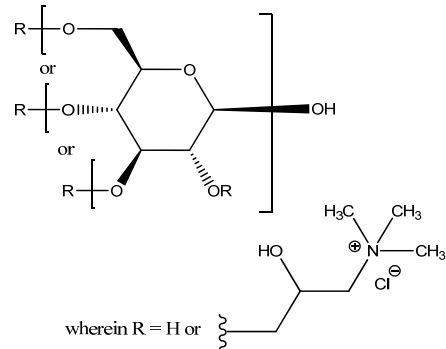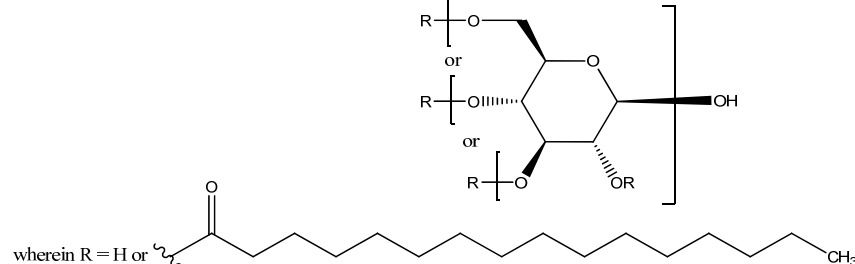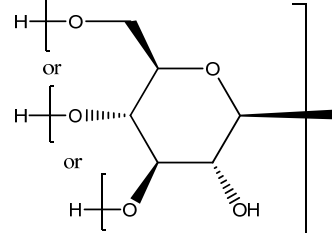
Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Dextran 9004-54-0	a high MW glucose polymer produced by the action of the bacteria, *Leuconostoc mesenteroides*, on a sucrose substrate[14]; the degree of branching is dependent on the source of dextrans and can vary from 0.5-60%[76]	binder; bulking agent	

wherein R = H or a branch of

| Carboxymethyl Dextran
9044-05-7 | the carboxymethyl derivative of dextran[14] | film former; viscosity increasing agent-aq. | |

wherein R = H, , or a branch of

| Dextran Hydroxypropyltrimonium Chloride
83855-79-2 | the quaternary ammonium compound formed by the reaction of dextran with glycidyltrimethylammonium chloride[14,77] | hair conditioning agent | |
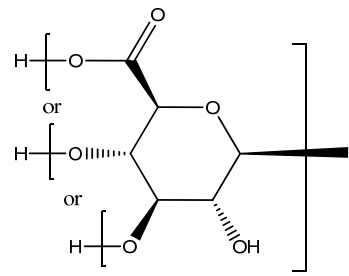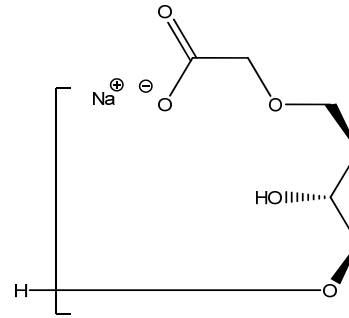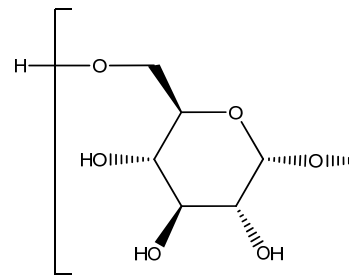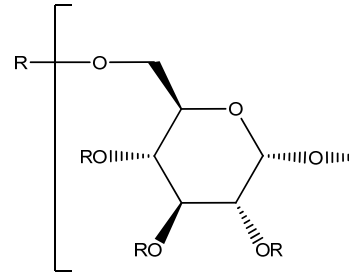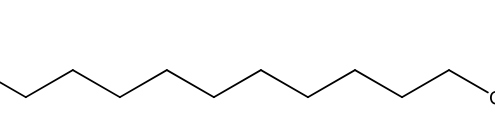
wherein R = H, , or a branch of

23

Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Sodium Carboxymethyl Dextran 39422-83-8	the sodium salt of a carboxymethyl dextran[14]	binder; emulsion stabilizer; viscosity increasing agent-aq.	

wherein R = H, Na, or a branch of

| Dextran Sulfate 9042-14-2 | the sulfuric acid ester of dextran[14] | binder; skin conditioning agent-misc. | |

wherein R = H, or a branch of

| Sodium Dextran Sulfate 9011-18-1 | the sodium salt of the sulfuric acid ester of dextran[14] | suspending agent-nonsurfactant | |

wherein R = H, Na, or a branch of
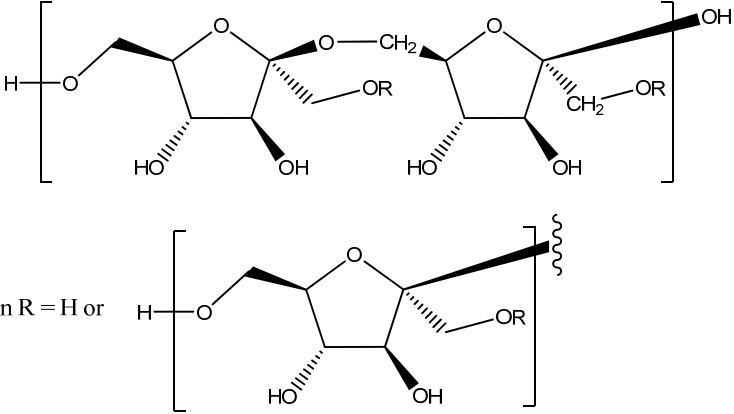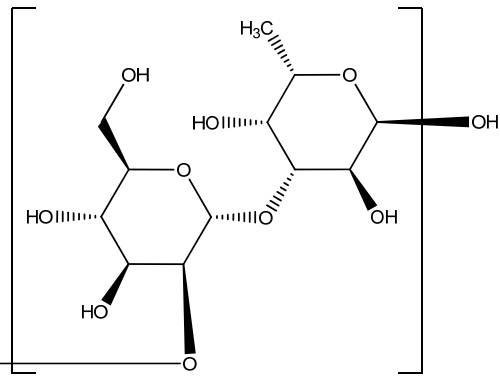
Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Beta-Glucan 55965-23-6 [CAS No. is specific to (1→3)(1→4)] 53238-80-5 [CAS No. is specific to (1→3)(1→6)]	a polysaccharide consisting of β(1-3), β(1-4), and β(1-6) linked glucose units[14]	bulking agent; skin conditioning agent-misc	

Beta-Glucan Hydroxypropyl-trimonium Chloride	the quaternary ammonium compound formed by the reaction of beta-glucan with glycidyltrimethylammonium chloride	anti-static agent; hair conditioning agent; skin conditioning agent - misc	

Beta-Glucan Palmitate	the product obtained by the condensation of beta-glucan with palmitoyl chloride[14]	skin conditioning agent - misc	

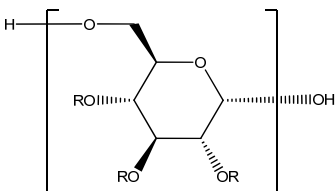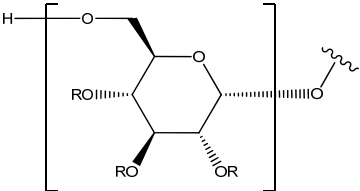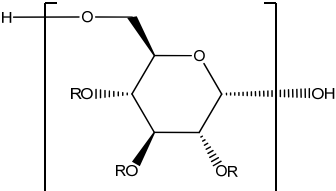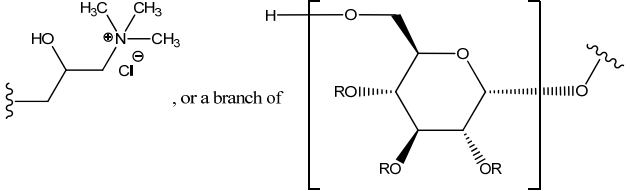|			
Hydrolyzed Beta-Glucan	the partial hydrolysate of beta-glucan, derived by acid, enzyme or other method of hydrolysis	skin conditioning agent - misc	

Table 1. Definition, Function, and Idealized Structure

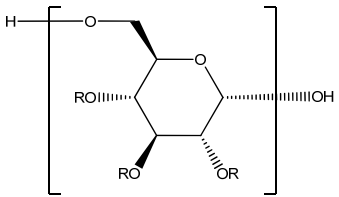| Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Oxidized Beta-Glucan	the product obtained by the oxidation of beta-glucan,[14] wherein the 6-position is oxidized to the acid[78]	anti-acne agent; anti-dandruff agent; anti-fungal agent; antimicrobial agent; deodorant agent; exfoliant; skin conditioning agent - misc	

Sodium Carboxymethyl Beta-Glucan 9050-93-5 [CAS No. specific to (1→3) β-D-Glucan]	the sodium salt of a carboxymethyl ether of beta-glucan, wherein the ether substitution occurs primarily at the 6-position[79]; 3 of 4 glucose-units of the β-1,3-glucopyranose are carboxymethylated[80]	binder; viscosity increasing agent - misc	

Pullulan 9057-02-7	an extracellular polysaccharide produced from starch by cultivating the yeast, *Aureobasidium pullulans*; composed of 1→6 linked maltitriose residues (a maltitriose is a block of three 1→4 linked, glucose resides)[81]	binder; film former	

Myristoyl Pullulan 1033464-89-9	the reaction product of myristoyl chloride and pullulan[14]	film former; hair fixative; hair-waving/straightening agent	

wherein R = H or
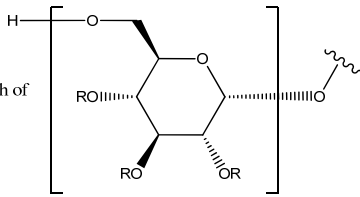
Table 1. Definition, Function, and Idealized Structure

Ingredient; CAS No.	Definition	Reported Function(s)[14]	Idealized Structure
Levan 9013-95-0	a polyfructose (fructofuranose), with some branching, produced by enzymatic action on sucrose[81]; the degree of branching varies by organism, but has been reported as high as 20%[82]	film former; skin protectant; viscosity increasing agent – aq.	

wherein R = H or

| Alcaligenes Polysaccharides
188846-47-1 [for the pure disaccharide repeat unit] | the polysaccharides produced by a bacterial culture of *Alcaligenes latus*; composed of mannose and fucose[83] | emulsion stabilizer; humectant; skin conditioning agent – humectant; viscosity increasing agent-aqueous | |

Table 2. Chemical and physical properties

Property	Description
Xanthan Gum	
appearance	cream-colored, odorless, free-flowing powder[35] white/beige powder with a characteristic odor[84]
MW	1,000,000 – 10,000,00 [33] varies within a very wide range[85]
solubility	dissolves readily in water with stirring to give highly viscous solutions at low concentrations[35] completely soluble in water, forming colloidal solution; insoluble in alcohol[84] readily soluble in hot or cold water to form neutral, viscous, and nonthixotropic solutions that have relatively high viscosity[33]
stability	resistant to heat degradation[35]
ionic nature	anionic[9,81]
pH	5.5-8.5 (1% solution, 25°C) [84]
Gellan Gum	
appearance	off-white powder[85-87]
types	native, deacetylated, clarified (i.e., filtered deacetylated gum)[86]
MW	varies within a very wide range[85] ~500,000[86] >70,000, with 95% above 500,000[39]
solubility	soluble in hot or cold deionized water[87] soluble in water; insoluble in ethanol[86]
viscosity	can exhibit high viscosity in solution[11] high-acyl gellan gum is viscous; deacylated gellan gum (treated with an alkali) has relatively low solution viscosity[13] cold dispersions of native gellan gum provide extremely high viscosities, and the solutions are highly thiotropic; the viscosity decreases with heating as the gum hydrates; hot native gum solutions are more viscous than deacylated gellan gum solutions[13]
gelling property	forms a weak gel in water in its native state, but forms a rigid gel after treatment with an alkali[12]
ionic nature	anionic[9,81]
hydration	native (acylated) gellan gum will swell in deionized water forming a very thick particulate system, and the hydration temperature is reached at ~70°C; the swelling behavior and hydration temperature is altered in the presence of ions[13] deacylated gum will only partially hydrate in cold deionized water, with hydration occurring with a heated deionized water temperature of ~70°C; also hydration is poor in mildly acidic conditions and in the presence of divalent ions[13]
Welan Gum	
MW	865,000-932,000[73]
viscosity	non-gelling polysaccharide forming highly viscous aq. solutions[88]
ionic nature	anionic[9]
Biosaccharide Gum-1	
MW	1,000,000 (avg)[89]
Biosaccharide Gum-4	
MW	2,000,000 (avg)[89]
Dextran	
appearance	Dextran 1: white to off-white powder[35]
MW	Dextran 1: ~1000 (avg) [90] Dextran 40: ~25,000 (solution has an avg MW of 40,000)[71] Dextran 70: ~40,000 (solution has an avg MW of 70,000)[71] MW distribution is dependent on the source of the dextran[76]
solubility	Dextran 1: very soluble in water; sparingly soluble in alcohol[35] degree of solubility decreases with an increase in the degree of branching; dextrans with >43% branching are insoluble[76]
ionic nature	nonionic[9,81]
stability	stable under mild acidic and basic conditions[76]
pH	Dextran 1: 4.5-7.0 (15% aq. solution) [90]
specific rotation	Dextran 1: between +148° and +164° at 20°, for an aq. solution[90] Dextran 40: between +195° and +203° [90] Dextran 70: between +195° and +203° [90]
Dextran Sulfate	
MW	5000-500,000 [10]

Table 2. Chemical and physical properties

Property	Description
	Sodium Dextran Sulfate
appearance	white powder[35]
MW	4000-50,000 [35]
solubility	freely soluble in water[35]
	Sclerotium Gum
solubility	disperses easily in water at room temperature; refined grades dissolve readily in hot and cold water[8]
ionic nature	nonionic[9,81]
	Beta-Glucan
appearance	white to pale yellow powder with a slight odor[91] white to nearly white powder (as curdlan)[87]
MW	~500,000 (native state)[92]
solubility	as curdlan: insoluble in water, alcohol, and most organic solvents; soluble in alkaline solutions[8]
ionic nature	nonionic[9]
	Oxidized Beta-Glucan
MW	continuum of ~30,000 to >70,000[92]
	Sodium Carboxymethyl Betaglucan
appearance	white/brown solid[93] amber, white granulate with a characteristic isopropyl alcoholic odor[80]
MW	~100,000[93] 4.23×10^5 (avg)[79]
degree of substitution	0.75 ±1[80,93] (as a 2% aq. solution) 0.2-0.3[79]
solubility	solubility up to 98%[79]
pH (2% aq. solution)	~7 [93]
	Pullulan
appearance	white to slightly yellowish powder; tasteless; odorless (food grade) [94] tasteless, odorless fine white powder[95]
MW	can vary considerably; a commercially available product has a molecular mass of 200,000 Da[31] 8000 - >2,000,000; approx. 200,000 (mean) [94]
solubility	highly soluble in cold or hot water; not soluble in organic solvents, except dimethylformamide or dimethyl sulfoxide (DMSO) [32,94]
stability	stable in aq. solution over a wide pH range; decomposes upon dry heating, carbonizing at 250-280°C [31]
viscosity	dissolves in water producing a stable viscous solution; does not gel; viscosity is proportional to MW[94] solutions are of relatively low viscosity; viscosity is stable to heating, changes in pH, and most metal ions[96]
ionic nature	nonionic[9,81]
pH	5.0-7.0 (food grade) [94]
refractive index	significant positive linear correlation of concentration and refractive index at 20 and 45°C [94]
	Levan
MW	up to several million Daltons; typically 2×10^6 to 10^8 [7] usually >0.5 million, and can be as high as 40,000,000 [82]
particle size	partially forms nanoparticles in water; 224.3 nm in water and 251.8 nm in ethanol[57]
solubility	highly soluble in water at room temperature[7] water soluble; does not swell in water[82]
viscosity	"exceptionally low" intrinsic viscosity[82]
ionic nature	nonionic[81]
	Rhizobian Gum
MW	1,500,000 (native molecule, in the fermentation broth)[75]
melting point	~60°C; gets lower after sterilization[97]
viscosity	with a 10 g/l solution, the viscosity decreases as the pH increases[75]
	Alcaligenes Polysaccharides
MW	64,000 [83]

Table 3. Constituents/Impurities

Ingredient	Constituents/Impurities
Xanthan Gum	nitrogenous constituents equal to approx.. 1% nitrogen by wt; approx.. half of the nitrogenous matter is proteinaceous and contains amino acid residues in the same proportions as other food-grade gums; the remainder is present as amino sugars, nucleic acids, and nucleotides[29]
	food-grade: contains D-glucose and D-mannose as the dominant hexose units, and D-glucuronic acid and pyruvic acid; NMT 2 mg/kg lead; NMT 0.075% ethanol and isopropyl alcohol, singly or combined[98]
	2.5-4.8% pyruvic acid, present as side chains[33]
Gellan Gum	usually contains a small amount of nitrogen-containing compounds as a result of the fermentation procedure[86]
	< 2 mg/kg lead; <3% nitrogen; ,750 mg/kg isopropyl alcohol[86]
	native: can contain 10% protein and 7% ash; deacetylated: can contain 17% protein and 8% ash[86]
	gellan gum is characterized by the polysaccharide content, percent of o-acetylated substitution, and protein content (including nucleic acid residues and other organic nitrogen sources)[39]
Dextran Sulfate	can contain polymers of various MWs and degrees of sulfation[41]
Beta-Glucan	≥85% β-1,3-1,6-glucan, sucrose, yeast extract, minerals, *Auerobasidium pullulans*[91]
	as curdlan: ≥90% carbohydrate; ≤10% water[8]
	as food grade curdlan – NMT 0.5 mg/kg lead; microbial limits, aerobic plate count NMT 1000 CFU/g; *Escherichia coli*, negative in 1 g[99]
Sodium Carboxymethyl Betaglucan	~90% sodium carboxymethyl betaglucan; <0.5% nitrogen (protein); <1.0% glycolic acid; <0.05% chloroacetic acid; ~10% volatile matter[80]
Pullulan	≥90% glucan on a dried basis; main impurities are mono-, di-, and oligosaccharides from the starting material[31]
	>90% pullulan; <10% mono-, di-,and oligosaccharides; <0.1 ppm lead; <2 ppm arsenic; <5 ppm heavy metals (food grade)[94]

Table 4. Methods of Manufacture/Organisms Used in Production

Ingredient	Method
Xanthan Gum	pure-culture fermentation of glucose or corn syrup from the bacterium *Xanthomonas campestris*; the polysaccharide is recovered by precipitation and purification with isopropyl alcohol, followed by drying and milling[33,60]
Gellan Gum	aerobic submerged fermentation using the bacterium *Pseudomonas elodea*; small seed fermentation is followed by pasteurization; the gum is recovered by precipitation with isopropyl alcohol, followed by drying and milling[100]
	pure culture fermentation of carbohydrates by *Pseudomonas elodea*, purified by recovery with isopropyl alcohol, dried, and milled[86]
	gellan gum can be recovered using alcohol precipitation (high acyl gum) or with alkali (deacylated gum)[13]
	produced by an organism that appears to belong to the *Auromonas* (ATCC 31461) genus; glycerate substitution predominates over acetate[11]
	Sphingomonas paucimobilis produce gellan gum[86]
	production is affected by media components, carbon source, nitrogen source, precursors, agitation rate, pH, temperature, and oxygen[86]
Welan Gum	fermentative production by *Alcaligenes* CGMCC2428 [73]
	produced by an *Alcaligenes* species (ATCC 31555)[11,88]
Dextran	Dextran 40; Dextran 70: obtained by the controlled hydrolysis and fractionation of polysaccharides elaborated by the fermentative action of certain appropriate strains of *Leuconostoc mesenteroides* on a sucrose substrate[90]
	the fermentation process for reaching high MW dextran takes place at 25°C; at lower temperatures, the amount of low MW dextran increases; at >25°C, higher branching occurs[10]
	different strains of the same bacterium produce dextrans with differing branched structures[71]
	the sucrose concentration also affects branching; increased sucrose content the degree of branching and the yield of high MW dextran decreases[10]
	dextran can be synthesized by dextrinase of different *Gluconobacter* species[10]
	dextran can be produced enzymatically using cell-free culture supernatants that contain dextransucranase[10]
	dextran can be synthesized chemically via a cationic ring-opening polymerization of levoglucosan[10]
Carboxymethyl Dextran	carboxymethylation of dextran in water/organic solvent mixtures using monochloroacetic acid under strong alkaline conditions[10]
Sclerotium Gum	produced by *Sclerotium glucanicum*[81] or *Sclerotium rolfsii*[8]
Beta-Glucan	extraction of extracellular β-glucan produced by the black yeast *Aureobasidium pullulans*[91]
	produced by fungi, yeasts, and grains (such as oat and barley); beta-glucans present in cereal bran are commonly produced as agricultural by-products[6]
	as curdlan: pure-culture fermentation of a carbohydrate by a nonpathogenic and nontoxigenic strain of *Agrobacterium biobar* 1 (formerly *Alcaligenes faecalis* var. myxogenes) or *Agrobacterium radiobacter*[99]

Table 4. Methods of Manufacture/Organisms Used in Production

Ingredient	Method
Oxidized Beta-Glucan	oxidation of beta-glucan is performed using phosphoric acid and sodium nitrite, with the actual oxidant being NO_2 gas; extent of oxidation was typically 10-20%[92]
Sodium Carboxymethyl Betaglucan	derived from the inner cell walls for baker's yeast (*Saccharomyces cerevisiae*); 3 of 4 glucose units of the β 1,3-glucopyranose are carboxymethylated[80,93]
	particulate glucan and sodium hydroxide are mixed, the sodium salt of monochloroacetic acid in 95% ethanol is added and stirred, excess sodium hydroxide is neutralized, the product is washed with 80% ethanol and dried[79]
Pullulan	fermentation of liquefied corn starch by *Aureobasidium pullulans*; the fungal biomass is removed by microfiltration, the filtrate is heat-sterilized, and the pigments and other impurities are removed by adsorption and ion-exchange chromatography[31]
	one company reports the following raw materials: ammonium sulfate; beer yeast extract; calcium hydroxide; caustic soda; corn syrup; diatomaceous earth; diammonium phosphate; dipotassium phosphate; hydrochloric acid; ion exchange resin; magnesium sulfate; salts; silicone oil; sodium glutamate; zinc carbon chloride[94]
	produced by *Pullularia pullulans* IFO 6353, *Dermatium pullulans* IFO 4464, etc in a culture medium containing a carbon source (such as glucose, fructose, etc) under anaerobic conditions[101]
Levan	produced extracellularly from sucrose- and raffinose-based substrates by levansucrase from a wide range of taxa, such as bacteria, yeasts, and fungi, but mainly by bacteria[7]
	fermentation of *Zymomonas mobilis* in a medium that contains 10% sucrose and 1% yeast extract, centrifugation via ultrafiltration, precipitation by the addition of ethanol, resuspension with distilled water, and drying[57]
	sources include Erwinia herbicola, Aerobacter lavanicum, Streptococcus salivarius, Pseudomonas prunicola, Arthrobacter acetigenum, Bacillus polymyxa, Bacilus subtilis, Actinomyces sp.[81]
Rhizobian Gum	produced by fermentation of Rhizobium sp. strain[75]
	bacterial strain YAS 34 is isolated from the rhizosphere of the sunflower plant; selection of the isolates is carried out on high carbon:nitrogen ratio liquid media; the culture broth was inoculated and fermented; the exopolysaccharide is recovered with a multi-step downstream processing that includes heating and centrifugation; diafiltration is used to eliminate fermentation residue, followed by further purification by alcoholic precipitation[102]
Alcaligenes Polysaccharides	neutral polysaccharide: culture broth was precipitated with ethanol and redissolved in hot water, filtration was used to remove the water-insoluble cells and acidic polymers, and the polysaccharide was recovered by ethanol precipitation and further purified[83]

Table 5. Frequency and concentration of use according to duration and type of exposure

	# of Uses[15]	Max. Concs. of Use (%)[16]	# of Uses[15]	Max. Concs. of Use (%)[16]	# of Uses[15]	Max. Concs. of Use (%)[16]
	Xanthan Gum		**Dehydroxanthan Gum**		**Xanthan Gum Crosspolymer**	
Totals*	**3470**	**0.00001-6**	**15**	**0.1-1**	**2**	**0.03-5**
Duration of Use						
Leave-On	*2782*	*0.001-6*	*6*	*0.1-1*	*2*	*0.03-5*
Rinse-Off	*678*	*0.000001-6*	*9*	*0.4-0.8*	*NR*	*NR*
Diluted for (Bath) Use	*10*	*0.5-3*	*NR*	*NR*	*NR*	*NR*
Exposure Type						
Eye Area	292	0.001-2	1	NR	NR	NR
Incidental Ingestion	35	0.03-2	NR	NR	NR	NR
Incidental Inhalation-Spray	121[a]	0.2-1[a]; 0.05[b]	1[a]	0.1[a]-0.2	NR	NR
Incidental Inhalation-Powder	19	0.3-6	NR	NR	NR	NR
Dermal Contact	3179	0.001-6	12	0.1-0.8	2	0.03-5
Deodorant (underarm)	1[c]	0.005-0.6[c] not a spray: 0.4-1	NR	NR	NR	NR
Hair - Non-Coloring	129	0.000001-4	3	0.7-1	NR	NR
Hair-Coloring	59	0.2-6	NR	NR	NR	NR
Nail	11	0.2-3	NR	NR	NR	NR
Mucous Membrane	206	0.03-4	5	0.4	NR	NR
Baby Products	29	0.2-0.6	NR	NR	NR	NR

	# of Uses	Max. Concs. of Use (%)	# of Uses	Max. Concs. of Use (%)	# of Uses	Max. Concs. of Use (%)
	Gellan Gum		**Biosaccharide Gum-1**		**Biosaccharide Gum-2**	
Totals*	**37**	**0.0004-0.5**	**346**	**0.002-6**	**14**	**1**
Duration of Use						
Leave-On	*35*	*0.0004-0.5*	*301*	*0.002-6*	*10*	*1*
Rinse Off	*2*	*NR*	*43*	*0.006-5*	*4*	*NR*
Diluted for (Bath) Use	*NR*	*NR*	*2*	*NR*	*NR*	*NR*
Exposure Type						
Eye Area	5	0.0004	28	0.01-1	2	NR
Incidental Ingestion	1	0.0004	NR	0.08	NR	NR
Incidental Inhalation-Spray	NR	NR	3[a]	0.002[a]	NR	NR
Incidental Inhalation-Powder	6	0.0004	1	NR	NR	NR
Dermal Contact	34	0.0004-0.3	326	0.002-6	14	1
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair - Non-Coloring	1	0.5	19	NR	NR	NR
Hair-Coloring	NR	NR	1	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous Membrane	2	0.0004	4	0.08	NR	NR
Baby Products	NR	NR	NR	NR	NR	NR

	# of Uses	Max. Concs. of Use (%)	# of Uses	Max. Concs. of Use (%)	# of Uses	Max. Concs. of Use (%)
	Biosaccharide Gum-4		**Dextran**		**Sodium Carboxymethyl Dextran**	
Totals*	**48**	**0.00001-5**	**51**	**0.000005-0.2**	**10**	**NR**
Duration of Use						
Leave-On	*43*	*0.004-5*	*48*	*0.000005-0.1*	*10*	*NR*
Rinse-Off	*5*	*0.00001-0.006*	*3*	*NR*	*NR*	*NR*
Diluted for (Bath) Use	*NR*	*NR*	*NR*	*0.2*	*NR*	*NR*
Exposure Type						
Eye Area	7	0.00001-0.2	4	NR	NR	NR
Incidental Ingestion	NR	NR	NR	NR	NR	NR
Incidental Inhalation-Spray	NR	1[a]	1[a]	0.01[a]	NR	NR
Incidental Inhalation-Powder	NR	NR	NR	NR	NR	NR
Dermal Contact	48	0.00001-5	48	0.000005-0.2	10	NR
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair - Non-Coloring	NR	NR	NR	NR	NR	NR
Hair-Coloring	NR	NR	NR	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous Membrane	NR	NR	NR	0.2	NR	NR
Baby Products	NR	NR	NR	NR	NR	NR

Table 5. Frequency and concentration of use according to duration and type of exposure

	# of Uses[15]	Max. Concs. of Use (%)[16]	# of Uses[15]	Max. Concs. of Use (%)[16]	# of Uses[15]	Max. Concs. of Use (%)[16]
	Dextran Sulfate		**Sodium Dextran Sulfate**		**Sclerotium Gum**	
Totals	**9**	**0.01-0.1**	**9**	**0.005-0.5**	**193**	**0.003-2**
Duration of Use						
Leave-On	*9*	*0.01-0.1*	*9*	*0.005-0.5*	*155*	*0.003-2*
Rinse Off	*NR*	*NR*	*NR*	*NR*	*37*	*0.003-1*
Diluted for (Bath) Use	*NR*	*NR*	*NR*	*NR*	*1*	*0.003*
Exposure Type						
Eye Area	9	NR	2	0.01-0.2	25	0.2-0.8
Incidental Ingestion	NR	NR	NR	NR	NR	NR
Incidental Inhalation-Spray	NR	NR	NR	NR	13[a]	0.003
Incidental Inhalation-Powder	NR	NR	NR	NR	3	NR
Dermal Contact	9	0.01-0.1	9	0.005-0.5	168	0.003-2
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair - Non-Coloring	NR	NR	NR	NR	20	0.003-1
Hair-Coloring	NR	NR	NR	NR	NR	0.8
Nail	NR	NR	NR	NR	1	0.6
Mucous Membrane	NR	NR	NR	NR	12	0.003-0.5
Baby Products	NR	NR	NR	NR	3	NR

	Hydrolyzed Sclerotium Gum		**Beta-Glucan**		**Sodium Carboxymethyl Beta-Glucan**	
Totals*	**NR**	**1**	**137**	**0.000001-0.1**	**67**	**0.0001-0.1**
Duration of Use						
Leave-On	*NR*	*1*	*103*	*0.0002-0.1*	*56*	*0.0002-0.1*
Rinse-Off	*NR*	*NR*	*34*	*0.000001-0.03*	*11*	*0.0001-0.1*
Diluted for (Bath) Use	*NR*	*NR*	*NR*	*NR*	*NR*	*NR*
Exposure Type						
Eye Area	NR	NR	9	NR	6	0.04
Incidental Ingestion	NR	NR	2	NR	NR	NR
Incidental Inhalation-Spray	NR	NR	NR	NR	NR	NR
Incidental Inhalation-Powder	NR	NR	7	NR	NR	0.02
Dermal Contact	NR	1	128	0.0002-0.1	66	0.0001-0.1
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair - Non-Coloring	NR	NR	4	0.000001	NR	0.04
Hair-Coloring	NR	NR	NR	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous Membrane	NR	NR	8	0.03	NR	0.1
Baby Products	NR	NR	12	NR	NR	0.02

	Pullulan		**Rhizobian Gum**		**Hydrolyzed Rhizobian Gum**	
Totals*	**45[d]**	**0.03-17[e]**	**5**	**NR**	**4**	**0.4-3**
Duration of Use						
Leave-On	*38*	*0.2-12*	*4*	*NR*	*3*	*0.4-3*
Rinse-Off	*1*	*0.03*	*1*	*NR*	*1*	*NR*
Diluted for (Bath) Use	*NR*	*NR*	*NR*	*NR*	*NR*	*NR*
Exposure Type						
Eye Area	10	3	2	NR	2	3
Incidental Ingestion	6	17[e]	NR	NR	NR	NR
Incidental Inhalation-Spray	1[a]	NR	NR	NR	NR	NR
Incidental Inhalation-Powder	NR	NR	NR	NR	NR	NR
Dermal Contact	36	0.2-3	5	NR	4	0.4-3
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair - Non-Coloring	NR	12	NR	NR	NR	NR
Hair-Coloring	NR	0.03	NR	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous Membrane	6	17[d]	NR	NR	NR	NR
Baby Products	NR	NR	NR	NR	NR	NR

Table 5. Frequency and concentration of use according to duration and type of exposure

	# of Uses[15]	Max. Concs. of Use (%)[16]	# of Uses[15]	Max. Concs. of Use (%)[16]	# of Uses[15]	Max. Concs. of Use (%)[16]
	Alcaligenes Polysaccharides					
Totals*	**15**	0.005-0.3				
Duration of Use						
Leave-On	*13*	*0.3*				
Rinse-Off	*2*	*0.005*				
Diluted for (Bath) Use	*NR*	*NR*				
Exposure Type						
Eye Area	2	NR				
Incidental Ingestion	NR	NR				
Incidental Inhalation-Spray	NR	NR				
Incidental Inhalation-Powder	NR	NR				
Dermal Contact	15	0.005-0.3				
Deodorant (underarm)	NR	NR				
Hair - Non-Coloring	NR	NR				
Hair-Coloring	NR	NR				
Nail	NR	NR				
Mucous Membrane	NR	NR				
Baby Products	NR	NR				

* Because each ingredient may be used in cosmetics with multiple exposure types, the sum of all exposure types my not equal the sum of total uses.

NR – no reported uses

[a] includes suntan products, in that it is not known whether or not the reported product is a spray

[b] this product is a pump spray

[c] it is not known whether or not the product is a spray

[d] the total of uses does not equal the duration of use for leave-on, rinse-off, and diluted for bath use because 6 uses are in oral hygiene products

[e] the use at 17% is in a breath freshener that dissolves in the mouth

Table 6. Ingredients Not Reported to be Used

Hydroxypropyl Xanthan Gum	Dextran Hydroxypropyltrimonium Chloride
Undecylenoyl Xanthan Gum	Beta-Glucan Hydroxypropyltrimonium Chloride
Xanthan Hydroxypropyltrimonium Chloride	Beta-Glucan Palmitate
Welan Gum	Hydrolyzed Beta-Glucan
Biosaccharide Gum-3	Oxidized Beta-Glucan
Biosaccharide Gum-5	Myristoyl Pullulan
Pseudoalteromonas Exopolysaccharides	Levan
Carboxymethyl Dextran	

Table 7. Examples of Non-Cosmetic Uses

Ingredient	Non-Cosmetic Use
Xanthan Gum	direct food additive[24]; indirect food additive[26]; stabilizer, thickener, and emulsifying agent in water-based pharmaceutical preparations[33]; used in oil and gas drilling and completion fluids[35]; stabilizer in the agrochemical industry and in water based paints and water-based printing inks, and other industrial uses.[103]
Gellan Gum	direct food additive[25]; thickener and gelling agent in the food industry[104]; novel drug-delivery vehicle, film formation for transdermal drug delivery, component in controlled-release systems[13]; alternative to agar for microbiological media[11,104,105] and plant tissue culture[104]
Welan Gum	thermostable thickener for industrial and oilfield application[11]; suspending, stabilizing, emulsifying, and thickening agent for food, coating materials, medicine, concrete additives, and enhanced oil recovery[73]
Dextran	GRAS indirect food additive[28]; approved active ingredient for OTC use (as dextran 70) as an ophthalmic demulcent at 0.1% when used with another approved polymeric demulcent[34]; a plasma volume expander (as dextran 70) and as a blood flow adjuvant (as dextran 40)[35]; 99mTc-labeled dextran is used as a tracer in lymphoscintigraphy[49]
Sodium Dextran Sulfate	clinical reagent[35]
Sclerotium Gum	pharmaceutical applications include use in table coatings, ophthalmic solutions, and injectable antibiotic solutions; thickening in the oil industry; drilling's mud and enhanced oil recovery; preparation of adhesives, water colors, printing inks; preparation of liquid animal feed[8]
Beta-Glucan	direct food additive (as curdlan)[27]
Pullulan	glazing agent, as a film-forming agent, as a thickener, and as a carrier in the production of capsules for dietary supplements, coatings for coated tablets, and edible flavored films[31]; can be used in wound-healing compositions; denture adhesive; photographic, lithographic, and electronic applications[96]
Levan	agricultural applications[7]

Table 8. Acute toxicity studies

Ingredient	Animals	No./Group	Vehicle	Concentration/Dose/Protocol	LD$_{50}$/Results	Reference
				ORAL		
Xanthan Gum	mice	not specified	water	not specified	>1 g/kg	[106]
Xanthan Gum	rats	not specified	not specified	5 g/kg, max	>5 g/kg	[107]
Xanthan Gum	dogs	not specified	not specified	20 g/kg, max	>20 g/kg	[107]
Gellan Gum	rats, M/F	not specified	not specified	administered in diet and by gavage; non-acetylated; >95% polysaccharide	>5 g/kg	[39]
Beta-Glucan (as curdlan)	mice, M/F	not specified	water	tested as a 10% suspension	>10 g/kg	[42]
Beta-Glucan (as curdlan)	rats, M	not specified	water	tested as a 10% suspension	>10 g/kg	[42]
Beta-Glucan (highly pure extract of *S. cerevisiae*)	rats	5M/5F	water	100 mg/ml suspension administered at 2 g/kg bw (20 ml/kg bw) by gavage	>2 g/kg	[108]
Sodium Carboxymethyl Beta-Glucan (>90% pure)	ddy mice	6M/6F	purified water	2 g/kg of a 20 aq. solution	>2 g/kg; no signs of toxicity	[109]
Pullulan	mice	not specified	olive oil	not specified	>14 g/kg	[110]
				INHALATION		
Xanthan Gum	albino rabbits	5	none	calculated chamber conc of 21 mg/l; 1 h exposure	>21 mg/l; no toxicity; no gross lesions at necropsy	[38]
Gellan Gum	rats, M/F	not specified	not specified	non-acetylated; >95% polysaccharide; duration of exposure not stated	>5.09 mg/l	[39]
Beta-Glucan (as curdlan)	guinea pigs	5	not specified	100 µg/ml curdlan for 4 h; continuous flow aerosol	no inflammatory cell response	[53]
Beta-Glucan (as curdlan)	guinea pigs	3	NaOH	1 µg/ml; continuous flow for 4 h	decrease in inflammatory cells with curdlan alone, increase in cells, especially neutrophils with NaOH	[54]
Beta-Glucan (as curdlan)	guinea pigs	5	not specified	6 µg/ml for 40 min, and animals were examined 4 or 24 h after exposure, orr 1 µg/ml for 4 h and animals were examined 5 or 9 days after exposure	in lung lavage: decrease in lymphocytes at 24 h in lung wall: statistically significant decrease in macrophages after 5 days; decrease in lymphocytes for 1-7 days	[54]
Beta-Glucan (as curdlan)	guinea pigs	5	not specified	100 µg/ml; continuous flow for 40 min or 4 h	in in lung lavage: slight decrease in macrophages; (almost significant) decrease lymphocytes	[54]
Beta-Glucan (as curdlan); MMAD 5 µm	guinea pigs	not specified	dust	office dust spiked with 1% beta-glucan (as curdlan; 280 g office dust was spiked with 2.8 g curdlan) for 4 h in a whole-body exposure chamber; necropsied 5 or 18 h after exposure	glucan-spiked dust produced a delayed subacute nasal congestion in guinea pigs compared to dust without beta-glucan; at 18 h after exposure, there was a significant decrease in in nasal volume	[52]
Beta-Glucan (as curdlan)	**humans**	36	dust	addition of 10 mg beta-glucan (as curdlan) to office dust (10 mg curdlan/g dust); subjects were exposed to the dust in a climate chamber for 180 min	compared to "clean" dust, nasal volume decreased, swelling in the nasal turbinate increased, and nasal eosinophil cell concentration increased	[55]
Pullulan	guinea pigs	5	not specified	100 µg/ml curdlan for 4 h; continuous flow aerosol;	no inflammatory cell response	[53]
				PARENTERAL		
Dextran Sulfate, avg MW of 7500, 47,000, and 458,000	mice and rats	not specified	not specified	0.1-2 g/kg, i.v. and i.p.	"some animals" died immediately; the largest MW dextran was more toxic	[111]
Beta-Glucan (as curdlan)	mice. M/F	not specified	saline	5% suspension, i.p.	males: 2.75 g/kg females: 2.5 g/kg	[42]
Beta-Glucan (as curdlan)	rats, M	not specified	saline	5% suspension, i.p.	2.75 g/kg; adhesions of beta-glucan to liver and spleen	[42]

Table 8. Acute toxicity studies

Ingredient	Animals	No./Group	Vehicle	Concentration/Dose/Protocol	LD$_{50}$/Results	Reference
Beta-Glucan (soluble; extract of *S. cerevisiae*)	ICR/HSD mice	not specified	not specified	≤1 g/kg, i.v.	>1 g/kg	[112]
Beta-Glucan (soluble; extract of *S. cerevisiae*)	Sprague-Dawley rats	not specified	not specified	0.5 g/kg, i.v.	>0.5 g/kg	[112]

Table 9. Repeated Dose Toxicity Studies

Ingredient	Animals/Group	Study Duration	Vehicle	Dose/Concentration	Results	Reference
				ORAL		
Xanthan Gum	rats (#/group not stated)	18 days	in diet	7.5%	paired feeding study – wt gains were similar to controls	[106]
Xanthan Gum	25 rats	30 days	in diet	1.5%	avg. wt of test animals was greater than controls; no gross lesions	[58]
Xanthan Gum	albino rats, 5M	91 days	in diet	3, 6, or 15%	reduced weight gain in the 15% group; no effect on organ wts; no microscopic lesions at necropsy in the 15% group	[106]
Xanthan Gum	rats, 5M/5F	110 days	in diet	0, 2.5, 5.0, or 10%	(the gum product consisted of drum-dried whole fermentation medium (beer)); no significant toxic effects	[38]
Xanthan Gum	CD rats, 30M/30F	2 yr	in diet	0, 0.25, 0.50, or 1.0 g/kg bw/day	no significant differences in growth rate, survival, hematology and clinical chemistry parameters, or organ weight were observed between treated and control animals; while not statistically significant (stat. sig.), there was an increase in uterine polyps in the high dose groups compared to controls, 5 in the high dose animals vs. 2 in controls	[61]
Xanthan Gum	beagle dogs, 3M/3F	12 wks	in diet	0, 0.25, or 0.5 g/kg bw/day	growth of high-dose males was slightly less than controls; the no-observable adverse effect level (NOAEL) was 0.25 g/kg bw/day	[38]
Xanthan Gum	beagle dogs, 2M/2F	12 wks	in diet	0, 1, or 2 g/kg bw/day; positive controls were given 20 g/kg bw/day powdered cellulose	immediate and persistent diarrhea in the 2 g/kg group; body wt loss was observed in treated and control animals, with the wt loss greatest in the 2 g/kg group; red blood cell counts, hemoglobin, and serum cholesterol levels were decreased in high dose animals; adrenal glands were slightly enlarged in the 2 g/kg group; no treatment-related microscopic lesion were observed at necropsy	[113]
Xanthan Gum	beagle dogs, 4M/4F	107 wks	in diet	0, 0.25, 0.37, and 1.0 g/kg/day	no significant differences in survival, body weight gain, feed consumption, organ weights, hematology parameters, or gross or microscopic lesions were observed between treated and control animals; a dose-related increased in urinary specific gravity was observed; ophthalmic findings were not considered treatment-related	[61]
Gellan Gum	20 rats	28 days	in diet	5%	no changes in urinalysis values; no gross lesions at necropsy	[114]
Gellan Gum; non-acetylated; >95% polysaccharide	Sprague-Dawley rats, 20M/20F	13 wks	in diet	0-6%	no mortality; no signs of toxicity	[39]
Gellan Gum; varied degree of acetylation; 58.5% polysac.	beagle dogs, 5M/5F	52 wks	in diet	0, 3, 4.5, or 6%	no mortality; feed consumption was greater in treated animals compared to controls; no adverse effects were observed	[39]
Gellan Gum; varied degree of acetylation; 58.5% polysac.	rhesus monkey, 2M/2F	28 days	not provided	0, 1, 2, or 3 g/kg, by gavage	no signs of toxicity	[39]
Dextran	albino rats, 6M	62 days	in diet	15%	wt gain was normal	[106]
Beta-Glucan (as curdlan)	CD-1 mice, 10M/10F	8 wk	in diet	0, 1, 5, 10, 20, or 30%; equiv to 0, 1.4, 7.1, 14, 29, and 43 g/kg bw, respectively	one female of the 30% group died; body wt gains of male mice of the 30% group were decreased compared to controls; no gross abnormalities; differences in stools and cecal wts were reported; the NOEL was 5% based on an increase in full cecal wts and large stools at higher doses	[42]
Beta-Glucan (as curdlan)	CD-1 mice, 100M/100F	99-114 wks (until survival was 20%)	in diet	0, 1, 5, or 15%	no treatment-related differences in survival or body wts; feed consumption was decreased by ~13% in 15% females; NOEL of 5% due to decreased feed consumption	[42]

Table 9. Repeated Dose Toxicity Studies

Ingredient	Animals/Group	Study Duration	Vehicle	Dose/Concentration	Results	Reference
Beta-Glucan (as curdlan)	Sprague-Dawley Ta rats; 5M	4 wks	in diet	0, 3, 10, or 30%; equiv to 0, 2.5, 8.5,or 30 g/kg bw 30% powder agar group	no difference in body wts; dose-related increase in neutrophils, with a stat. sig. increase in the 10% group; occult blood was present in the urine 4 10% rats, 1 30% rat, and 1 fed agar; relative (rel.) kidney wts were statistically significantly decreased in all groups; rel. liver weights were statistically significantly decreased and pituitary wts were stat. sig. increased in the 30% group	[42]
Beta-Glucan (as curdlan)	20 Sprague-Dawley rats, males	8 wks	in diet	0, 1, 5, or 15%	no mortality or signs of toxicity; body wt of animals of the 15% dose group were stat. sig. decreased; feed consumption in this group was slightly decreased	[42]
Beta-Glucan (as curdlan)	Sprague-Dawley rats, 10M/10F	3 mos	in diet	0, 5, 10, or 20%; equiv to 0, 4.4, 9, and 19 g/kg bw (males) and 0, 5.5, 12, and 24 g/kg bw (females)	body wt gains decreased with increasing dose, being stat. sig. in the 20% group; a stat. sig. decrease in platelet count in males and in total protein and globulin concs. in males and females of the 10 and 20% groups; stat. sig. increase in body wt of males of the 10 and 20% group; stat. sig. decreases in absolute liver wts in 20% males, absolute (abs.) kidney wts of 10 and 20% males, abs. and rel. ovary wts in 20% fe-males, abs. and rel. pituitary wts of in all females of all doses; and rel. adrenal wts in 10 and 20% males; differences in stools and cecal wts were reported; the NOEL was 5% based on fecal changes, diarrhea, and cecal wts and large stools at higher doses	[42]
Beta-Glucan (as mushroom beta-glu-can from *Ganoder-ma lucidum*)	CD (SD) IGS rats, 12M,12F	3 mos	sterile water; by gavage	0, 500, 1000, or 2000 mg/kg bw (10 ml/kg dosing volume)	no test article-related adverse effects on mortality, toxicity; opthalmo-scopy, body wts or body wt gains, clinical chemistry, hematology, or urinalysis; statistically significant decreases in testes wt in males of the low dose group and heart wts in females of the high dose group were not considered test-article related; there were no treatment related gross or microscopic lesions, the NOAEL was 2000 mg/kg bw	[42]
Beta-Glucan (as curdlan)	CD rats; 60M/60F	2 yrs	in diet	0, 1, 5, or 15%	no changes in mortality, behavior, appearance, or ophthalmic parame-ters; wt gain was decreased by ~10%, which was not stat. sig., in the 15% group; feed consumption was also reduced; no microscopic lesions were found the NOEL was 5% based on decreased feed consumption, body wt gain and increased cecal wts	[42]
Beta-Glucan (as curdlan)	CD rats; 60M/60F, of the F_{1a} generation of a reproductive tox. study	124-127 wks (20% survival)	in diet	0, 1, 5, or 15%	no changes in mortality, behavior, appearance, or ophthalmic parame-ters; body wts of the 15% group were stat. sig. decreased, and body wts were stat. sig. decreased in 5% males until wk 65; feed consumption was decreased in the 15% group; no changes in hematology or urinary parameters, but some stat. sig. changes were reported in blood chemistry; stat. sig increase in gross and microscopic incidences of uterine polyps in the 15% groups – the incidences were 0/450, 3/50, 4/51, and 7/50 for the 0, 1, 5, and 15% groups, indicating that the polyps were possible treatment-related; the NOEL was 1% based on increased cecal wts, decreased body wts and feed consumption, and the incidence of polyps	[42]
Beta-Glucan (as curdlan)	beagle dogs, 4M/4F	52 wks	in diet	0, 1, 5, or 15%	one 15% male died at 37 wks (not dose related); no stat. sig. treatment-related changes, except for fecal changes and cecal wts; NOEL was 5%based on fecal changes and cecal wts	[42]

Table 9. Repeated Dose Toxicity Studies

Ingredient	Animals/Group	Study Duration	Vehicle	Dose/Concentration	Results	Reference
Beta-Glucan (highly pure extract of *S. cerevisiae*)	SPF Fischer rats, 10M/10F	91 days	water	0, 2, 33.3, or 100 mg/kg bw/day, volume 0.5 ml/100 g bw, by gavage	no mortality; no stat. sig. differences in wt gains, feed consumption, or gross or microscopic lesions; a dose-dependent and stat. sig. increase in clotting time in males, isolated stat. sig. changes in some clinical chemistry parameters, and slight but stat. sig. incr. in absolute liver, kidney, heart, spleen, adrenal, and testicle wts in males and absolute kidney and thymus wts in females were not considered toxicologically significant; NOAEL was 100 mg/kg bw/day	[108]
Beta-Glucan (high-purity extract barley beta-glucan)	SPF Wistar rats, 5M/5F	28 days	in diet	0, 1, 5, or 10%, supplemented with ≤10% potato starch	no mortality; no sig. effects on body wts, feed consumption, or functional observational battery results; no treatment-related change in hematology or urinalysis values; empty caecum wt was increased	[115]
Beta-Glucan (extracted from *Candida albicans*)	Sprague-Dawley rats, 20M/20F	52 wks	sterile saline (using a rubber catheter)	0, 50, 100, or 200 mg/kg/day	no sig. effects on mortality, body wts, feed consumption, or hematology, clinical chemistry, or urinalysis parameters; with the exception of cecal enlargement with variable hyperplasia, not gross or microscopic lesions were noted; the NOEL was 100 mg/kg/day	[116]
Pullulan	8 Wistar rats, males (test and cellulose control groups)	4 or 9 wks	in diet	0, 5, 10, 20, or 40%; equiv. to 0, 2500, 5000, 10,000, or 20,000 mg/kg, respectively cellulose controls: 20 or 40% cellulose	body wt gains were decreased by day 10 in rats of the 20 and 40% groups when compared to untreated controls; wt gains of animals of the 5 and 10% pullulan group were lower than untreated controls after 7 wks, but this difference was not stat. sig.; similar decreases were observed in the animals fed cellulose; diarrhea was observed with 40% pullulan; rel. wts of the stomach, small intestine, and large intestine were increased in treated animals	[43,44]
Pullulan (200,000 MW)	SPF Wistar rats, 10M/10F	13 wks	in diet	0, 2.5, 5, or 10%,; equiv to 0, 1960, 4100, or 7900 mg/kg bw/day diet of the groups fed 0, 2.5, or 5% pullulan was supplemented with 10, 7.5, or 5% potato starch, respectively	no mortality; no dose-related clinical sign; stat. sig. reduced motor activity in females of the 5 and 10% groups was observed and appeared treatment related, but as a physiological phenomenon rather than a toxic effect; no difference in hematology parameters between treated and control groups; differences that were observed in clinical chem. parameters were not biologically significant or dose-related; dose-dependent, stat. sig. increases in abs. and rel. empty cecal wts were observed in males of the 5 and males and females of the 10% group; no microscopic changes were observed	[44]
Pullulan	Sprague-Dawley rats, 15M/15F	62 wks(was to be 24 mos; study was terminated because of poor survival due to pneumonia)	in diet	0, 1, 5, and 10%	no treatment-related effects on body wts, feed consumption or organ wts (except for cecal wts); changes in hematology or clinical chemistry parameters and microscopic lesions that were observed were not considered treatment-related; the NOAEL was 10% in the diet (equiv. to 4450 mg/kg bw/day)	[117]
INHALATION						
Beta-Glucan (as curdlan)	guinea pigs, 16F	5 wks	distilled water	100 µg/ml; 4 h/day, 5 days/wk	no significant change in lung lavage cells, but there was a decrease in the number of lymphocytes; slight but not statistically significant increase in macrophages and eosinophils in the lung wall cells; no lesions observed at microscopic examination of the lungs	[118]
Beta-Glucan (as curdlan)	guinea pigs, 6	5 wks	saline	100 µg/ml; 4 h/day, 5 days/wk; continuous flow exposure with a dose of 8 pg; animals were examined 24 h after last dose	the only effect on lung lavage cells was a slight decrease in neutrophils; there was no effect on the number of lung wall cells	[119]
PARENTERAL						
Xanthan Gum	mice	2 wks	water	5 mg in 0.5 ml water; 10 i.p. injections over 2 wks	no toxicity; there was no xanthan gum in the abdominal cavity at necropsy	[106]

Table 9. Repeated Dose Toxicity Studies

Ingredient	Animals/Group	Study Duration	Vehicle	Dose/Concentration	Results	Reference
Dextran, partly hy-drolysed, bacterial, 75,000 avg MW	16	103-113 wks	physiological saline	30 ml of 6% solution, i.v.	4 animals died and one was killed in moribund condtion; wt gain was similar to that of controls; increases in absolute heart and adrenal wts were probably statistically significant; statistically significant increase in liver and spleen wts; no microscopic lesions	[47]
Beta-Glucan (sol-uble; extract of *S. cerevisiae*)	mice	7 days	not specified	250 mg/kg bw, i.p.	no effect on weight gain	[112]
Beta-Glucan (sol-uble; extract of *S. cerevisiae*)	guinea pigs	7 days	not specified	250 mg/kg bw, i.p.	sig. 10% decrease in weight gain	[112]
Beta-Glucan (sol-uble; extract of *S. cerevisiae*)	ICH/HSD mice, M	30 days	not specified	≤1000 mg/kg bw, i.v.	sig. toxicity leading to hepatosplenomegaly in the 40 and 1000 mg/kg bw groups	[112]
Beta-Glucan (sol-uble; extract of *S. cerevisiae*)	ICH/HSD mice, M	60 days	not specified	≤1000 mg/kg bw, i.v.	dose-dependent increase in hepatosplenomegaly; was stat. sig. at 1000 mg/kg	[112]

Table 10. Genotoxicity studies

Ingredient	Concentration	Vehicle	Procedure	Test System	Results	Reference
IN VITRO						
Gellan Gum (non-acetyl-ated; >95% polysacc.)	10, 30, 100, 300, and 1000 μg/plate	not provided	Ames test, with and without metabolic activation	*Salmonella typhimurium* TA98, TA100, TA1537, TA1538, TA1535	negative	[39]
Gellan Gum (non-acetyl-ated; >95% polysacc.)	3, 5, 10, and 20 mg/ml	not provided	DNA repair test (details not provided)	rat hepatocytes	negative	[39]
Gellan Gum (non-acetyl-ated; >95% polysacc.)	3, 5, 10, and 20 mg/ml	not provided	V79/HGRPT (details not provided)	Chinese hamster lung fibroblasts	negative	[39]
Sodium Dextran Sulfate (54,000 MW)	1.0, 7.5, 25 mg/plate	not provided	Ames test; appropriate positive controls were used	S. typhimurium TA100, TA98	negative	[120]
Sodium Dextran Sulfate (54,000 MW)	0, 10, 100 μg	distilled water	hepatocyte primary culture (HPC)/DNA repair test; appropriate positive controls were used	rat hepatocytes	negative	[120]
Sodium Dextran Sulfate (54,000 MW)	0, 10, 100 μg	distilled water	intestinal mucosal cell (IMC)/DNA repair test; appropriate positive controls were used	intestinal mucosal cells from rat ileum or colon and rectum	negative	[120]
Beta-Glucan (as curdlan)	15-5000 μg/plate	sterile water	Ames test, with and without metabolic activation	*S. typhimurium* TA98, TA100, TA1537, TA1538, TA1535	negative	[42]
Beta-Glucan (as mush-room beta-glucan from *Ganoderma lucidum*)	313-5000μg/plate	not provided	Ames test, with and without metabolic activation	*S. typhimurium* TA98, TA100, TA102, TA1535, TA1537	negative	[121]
Beta-Glucan (as curdlan)	625-5000 μg/ml	tissue culture medium	chromosomal aberration assay, with and without metabolic activation	Chinese hamster ovary (CHO) cells	negative	[42]
Beta-Glucan (as mush-room beta-glucan from *Ganoderma lucidum*)	313-5000μg/ml	culture medium	chromosomal aberration assay, with and without metabolic activation	CHO cells	negative	[121]

Table 10. Genotoxicity studies

Ingredient	Concentration	Vehicle	Procedure	Test System	Results	Reference
Beta-Glucan (as curdlan)	12.5-5000 µg/ml	tissue culture medium	tk locus test, with and without metabolic activation	mouse lymphoma L518Y cells	negative	[42]
Sodium Carboxymethyl Beta-Glucan	0, 3-5000 µg/plate	Milli-Q water	Ames test, with and without metabolic activation, appropriate positive controls were used;	*S. typhimurium* TA98, TA100, TA1537, TA1535	negative	[122]
Sodium Carboxymethyl Beta-Glucan	0, 100-5000 µg/plate	Milli-Q water	Ames test, with and without metabolic activation; appropriate positive controls were used	*S. typhimurium* TA98, TA100, TA1537, TA1535	negative	[122]
Sodium Carboxymethyl Beta-Glucan	0, 195-50,000 µg/ml	purified water	Ames test, with and without metabolic activation; appropriate positive controls were used;	*S. typhimurium* TA98, TA100, TA1537, TA1535; *Escherichia coli* WP2 uvrA	negative	[123]
Sodium Carboxymethyl Beta-Glucan	0, 3125-50,000 µg/ml	purified water	Ames test, with and without metabolic activation; appropriate positive controls were used	*S. typhimurium* TA98, TA100, TA1537, TA1535; *Escherichia coli* WP2 uvrA	negative	[123]
Carboxymethyl-Glucan	12.5, 25, 50, 100, and 200 µg/ml		electrophoresis test in single-cell gel (comet assay)	CHO-k1 cells	negative	[124]
Pullulan	≤10,000 µg/plate	not provided	Ames test, with and without metabolic activation; appropriate positive controls were used	*S. typhimurium* TA98, TA100, TA1537, TA1535; *Escherichia coli* WP2 uvrA	negative	[117]
Pullulan	12 mg/ml	not provided	chromosomal aberration assay	Chinese hamster lung fibroblasts	negative	[125]
Pullulan	20 mg/plate	distilled water	rec assay; with and without metabolic activation	Bacillus subtilis	weak positive	[126]
IN VIVO						
Beta-Glucan (as curdlan)	0, 500, 1000, or 2000 mg/kg	water	micronucleus test; dosed by gavage at 24 h intervals; number of doses not specified; followed OECD Guideline No. 474	male and female CD-1 mice	negative	[42]
Beta-Glucan (as mushroom beta-glucan from *Ganoderma lucidum*)	0, 1250, 2500, and 5000 mg/kg bw	sterile water	micronucleus test; single dose by gavage ; blood samples were taken 24, 48, and 72 h after dosing	CD-1 mice, 5/sex/group	negative	[121]
Pullulan	1800 mg/kg bw.	not provided	micronucleus test; one i.p. injection	ddy mice	negative	[127]
Pullulan	4x1000 mg/kg bw	not provided	micronucleus test; four i.p. injections in 24 h	ddy mice	negative	[127]

Table 11. Dermal Irritation and Sensitization

Ingredient	Animals/Subjects/Gp	Dose/Conc/Vehicle	Procedure	Results	Reference
IRRITATION - NON-HUMAN					
Xanthan Gum	rats	1%; vehicle not provided	solution was applied daily for 15 days (details not provided)	not an irritant	[38]
Xanthan Gum	6 rabbits	0.5 ml of a 1% solution	primary cutaneous irritation test; occlusive patches on intact and abraded skin	non-irritant; primary irritation index (PII) – 0.13	[128]
Xanthan Gum	6 rabbits	0.5 ml of a 1% solution	primary cutaneous irritation test; occlusive patches on intact and abraded skin	non-irritant; PII – 0.13	[128]
Xanthan Gum	3 rabbits	2 ml/animal of a 1% solution	6-wk cumulative cutaneous irritation test, 5 applications/wk	very well tolerated; mean max. irritation index - 0	[128]
Xanthan Gum	3 rabbits	1%	6-wk cumulative cutaneous irritation test, 5 applications/wk	very well tolerated; MMII - 0	[128]
Xanthan Gum	rabbits	5% aq.	daily applications to shaved skin (details not provided)	localized irritation with bleeding and cracking; the effects may have been due to continuous moistening of the skin	[106]
Beta-Glucan	rabbits	not provided	primary skin irritation test (details not provided)	not an irritant	[91]
Beta-Glucan	rabbits	not provided	repeated skin irritancy test (details not provided)	not an irritant	[91]

Table 11. Dermal Irritation and Sensitization

Ingredient	Animals/Subjects/Gp	Dose/Conc/Vehicle	Procedure	Results	Reference
Sodium Carboxy-methyl Beta-Glucan (>90% pure)	Japanese white rabbits, 6M	0.5 g moistened with 0.2 ml distilled water	occlusive patches were applied to intact and abraded shaved skin for 24 h	non- to mildly irritating; primary irritation index (PII) of 0.33/8 at test site and 0.29/8 for adhesive control; no irritation was reported for intact skin	[129]
Sodium Carboxy-methyl Beta-Glucan (>90% pure)	3 guinea pigs	2, 10, or 50% (w/v) in distilled water	24-h occlusive patch test; this test was used to determine test concentration for the maximization study described below	slight skin irritation was observed at a concentration of 50%	[130]
IRRITATION - HUMAN					
Beta-Glucan	not provided	not provided	occlusive patch test	no an irritant	[91]
Sodium Carboxy-methyl Beta-Glucan (>90% pure)	40 subjects; 27M/13F	applied neat; small amount of vaseline was used for adhesion	24-h occlusive patch test; 0.1 g of test material was applied	not a primary skin irritant; no irritation was observed	[131]
SENSITIZATION – NON-HUMAN					
Xanthan Gum	guinea pigs, 18M	0.1%; vehicle not provided	intradermal challenge test; test solution was injected intracutaneously 3x/wk for 10 injections; the challenge was performed after a 10-day non-treatment period	not a sensitizer	[38]
Beta-Glucan	guinea pigs	not provided	skin sensitization test (details not provided)	not a sensitizer	[91]
Sodium Carboxy-methyl Beta-Glucan (>90% pure)	Hartley guinea pigs, 6F	10% in distilled water	maximization study using Freund's complete adjuvant and SLS; 0.1% aq. 2,4-dinitro-1-chlorobenzenze was used as the positive control dose volumes were 0.1 ml for intradermal induction, 0.2 ml for dermal induction, and 0.1 ml for challenge	not a sensitizer	[130]
SENSITIZATION - HUMAN					
Beta-Glucan (as Curdlan)	213 subjects; M	dose not provided; was an aq. paste	modified Draize 'multiple insult' patch test; occlusive patches were applied every other day for 10 applications; a 48-h challenge was performed after a 10-14 day non-treatment period	trace, insignificant, irritation observed during induction; not a sensitizer	[42]
Sodium Carboxy-methyl Beta-Glucan	32 subjects; 8M/24F	induction and challenge: 2% in distilled water	induction: 9 24-h occlusive patches were applied challenge: occlusive patch applied after a 10-14 day non-treatment period to a previously unpatched site reactions were graded on a scale of 0-4	not a sensitizer during induction, 2 subjects had doubtful reactions and one had a grade 1 reaction	[132]
PHOTOALLERGY- HUMAN					
Sodium Carboxy-methyl Beta-Glucan	8 male/24 female subjects	induction and challenge: 2% in distilled water	induction: 6 24-h patches were applied to non-tanned skin; the test site was irradiated with 2x MED UVB within 10 min after patch removal challenge: applied after a 10-14 day non-treatment period, a 24-h patch was applied to a previously unpatched site; the site was irradiated with 18 J/cm^2 UVA within 10 min after patch removal MULTITESTER light source was used; 0-4 scale used for scoring	not photoallergenic 31 subjects had grade 1 skin reactions, and on had a grade 2 reaction, to UVB exposure during induction 1 subject had a doubtful reaction after challenge	[132]

Table 12. Ocular Irritation

Ingredient	Animals/Subjects/Group	Concentration/Vehicle	Procedure	Results	Reference
			ALTERNATIVE STUDIES		
Sodium Carboxy-methyl Beta-Glucan		5% stock solution containing 2% Sodium Carboxymethyl Beta-Glucan	Ocular tolerance test using HET-CAM method	weakly irritant	[133]
			NON-HUMAN		
Xanthan Gum	rabbits	1%	ocular irritation test	non-irritating; acute ocular irritation index (AOII) - 2.50/110	[128]
Xanthan Gum	rabbits	1%	ocular irritation test	non-irritating; AOII – 5.83/110	[128]
Xanthan Gum	rabbits	1%	instilled in the conjunctival sac of rabbit eyes for 5 days (details were not provided)	not irritating	[38]
Gellan Gum	New Zealand White rabbits, 3M	0.2 or 0.3%	Draize study; 50 µl of the test solution was instilled into the conjunctival sac of the eye three times a day for 10 days	not irritating	[134]
Gellan Gum	albino rabbits	0.8%	details not provided	not irritating	[135]
Sodium Carboxy-methyl Beta-Glucan (>90% pure)	Japanese white rabbits, 3M	undiluted	0.1 mg were instilled into the conjunctival sac of one eye, and the contralateral eye served as an untreated control; the eyes of one group were rinsed 1 min after instillation	practically non-irritating; slight redness of the conjunctiva in both washed and unwashed eyes at 1 h, considered due to the powder	[136]
			HUMAN		
Gellan Gum	3 subjects	0.1-0.5%; vehicle not specified	25 µl of a gel formulation was instilled in the conjunctival sac of the eye, remained in contact with the eye for 9-52 min	not irritating	[134]

REFERENCES

1. Johnson WJ, Bergfeld WF, Belsito DV, Hill RA, Klaassen CD, Liebler DC, Marks JG, Shank RC, Slaga TJ, Snyder PW, and Andersen FA. Final report on the safety assessment of galactomannans as used in cosmetics. 2012. Available from the Cosmetic Ingredient Review. http://www.cir-safety.org.

2. Becker LC, Bergfeld WF, Belsito DV, Klaassen CD, Marks JG, Shank RC, Slaga TJ, Snyder PW, and Andersen FA. Final report of the safety assessment of hyaluronic acid, potassium hyaluronate, and sodium hyaluronate. *Int J Toxicol*. 2009;28:(Suppl 1):5-67.

3. Elder RL (ed). Final report on the safety assessment of tragacanth gum. *J Am Coll Toxicol*. 2011;6:(1):1-22.

4. Andersen FA (ed). Final report on the safety assessment of acacia catechu gum, acacia concinna fruit extract, acacia dealbata leaf extract, acacia dealbata leaf wax, acacia decurrens extract, acacia farnesiana extract, acacia farensiana flower wax, acacia farnesianan gum, acacia senegal extract, acacia senegal gum, and acacia senegal gum extract. *Int J Toxicol*. 2005;24:(Suppl 3):75-118.

5. Becker LC, Bergfeld WF, Belsito DV, Hill RA, Klaassen CD, Lin FSD, Marks JG, Shank RC, Slaga TJ, Snyder PW, and Andersen FA. Safety assessment of trimoniums as used in cosmetics. 2010. Available from the Cosmetic Ingredient Review. http://www.cir-safety.org.

6. Driscoll M, Hansen R, Ding C, Cramer DE, and Yan j. Therapetuic potential of various β-glucan sources in conjuction with anti-tumor monoclonal antibody in cancer therapy. *Cancer Biology & Therapy*. 2009;8:(3):218-225.

7. Rhee S, Song K, Kim C, and et al. Levan. Chapter: 14. In: *Biopolymers*. 2002:351-377.

8. Manjanna KM, Shivakumar B, and Pramodkumar TM. Natural exopolysaccharides as novel excipients in drug delivery: A review. *Archives of Applied Science Research*. 2009;1:(2):230-253.

9. Mathur V and Mathur NK. Microbial polysaccharides based food hydrocolloid additives. *Science Tech Entrepreneur*. 2006. pp.1-10.

10. Heinze T, Liebert T, Heublein B, and Hornig S. Functional polymers based on dextran. *Adv Polym Sci*. 2006;205:199-291.

11. Moorhouse R. Structure/property relationships of a family of microbial polysaccharides. Symposium on the Applications and Modifications of Industrial Polysaccharides. 1987.

12. Kuo M, Mort AJ, and Dell A. Identification and location of L-glycerate, an unusual acyl substituent in gellan gum. *Carbohydrate Research*. 1986;156:(November):173-187.

13. Valli RC and Miskiel FJ. Gellan gum. *Food Science and Technology (New York, NY, United States)*. 2001;113:(Handbook of Dietary Fiber):695-720.

14. Gottschalck TE and Breslawec HP. International Cosmetic Ingredient Dictionary and Handbook. 14 *ed*. Washington, DC: Personal Care Products Council, 2012.

15. Food and Drug Administration (FDA). Frequency of use of cosmetic ingredients. *FDA Database*. 2012. Washington, DC: FDA.Received from the FDA in May 2012.

16. Personal Care Products Council. 10-26-2011. Concentration of Use by FDA Product Category: Microbial Polysaccharides. Unpublished data submitted by Personal Care Products Council. 10 pages.

17. Personal Care Products Council. 3-28-2012. Concentration of Use by FDA Product Category: Xanthan Hydroxypropyltrimonium Chloride. Unpublished data submitted by Personal Care Products Council. 1 pages.

18. Personal Care Products Council. 7-9-2012. Comments on the Tentative Report of Microbial Polysaccharide Gums. Unpublished data submitted by Personal Care Products Council. 1 pages.

19. Johnsen MA. The influence of particle size. *Spray Technology and Marketing*. 2004;November:24-27.

20. Rothe H. Special Aspects of Cosmetic Spray Evaluation. 9-26-2011. Unpublished data presented at the 26 September CIR Expert Panel meeting. Washington, D.C.

21. Rothe H, Fautz R, Gerber E, Neumann L, Rettinger K, Schuh W, and Gronewold C. Special aspects of cosmetic spray safety evaluations: Principles on inhalation risk assessment. *Toxicol Lett*. 2011;205:(2):97-104.

22. Bremmer HJ, Prud'homme de Lodder LCH, and Engelen JGM. Cosmetics Fact Sheet: To assess the risks for the consumer; Updated version for ConsExpo 4. 2006. Report No. RIVM 320104001/2006. pp. 1-77.

23. European Commission. Health and Consumers Cosmetic Cosing Database. http://ec.europa.eu/consumers/cosmetics/cosing/. 2011. Date Accessed 10-20-2011.

24. Food and Drug Administration (FDA). 21CFR172.695. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=172.695&CFRPart=&FRSearch=. 4-1-2011. Date Accessed 10-21-2011.

25. Food and Drug Administration (FDA). 21CFR172.665. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=172.665&CFRPart=&FRSearch=. 4-1-2011. Date Accessed 10-21-2011.

26. Food and Drug Administration (FDA). 21CFR176.170. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=176.170&CFRPart=&FRSearch=. 4-1-2011. Date Accessed 10-21-2011.

27. Food and Drug Administration (FDA). 21CFR172.809. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=172.809. 4-1-2011. Date Accessed 11-19-2011.

28. Food and Drug Administration (FDA). 21CFR186.1275. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=186.1275&CFRPart=&FRSearch=. 4-1-2011.

29. Joint FAO/WHO Expert Committee on Food Additives (JEFCA). Evaluation of certain food additives and contaminants. Thirtieth report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series 751. http://whqlibdoc.who.int/trs/WHO_TRS_751.pdf. World Health Organization (WHO): Geneva. 1987. Date Accessed 3-22-2012.

30. Joint FAO/WHO Expert Committee on Food Additives (JEFCA). Evaluation of Certain Food Additives and Contaminants. WHO Technical Report Series 806. Thirty-seventh report of the Joint FAO/WHO Expert Committee on Food Additives. http://whqlibdoc.who.int/trs/WHO_TRS_806.pdf. World Health Organization: Geneva. 1991. Date Accessed 3-22-2012.

31. Joint FAO/WHO Expert Committee on Food Additives (JEFCA). Evaluation of certain food additives. WHO Technical Report Series 934. Sixty-fifth report of the joint FAO/WHO Committee on food additives. http://whqlibdoc.who.int/trs/WHO_TRS_934_eng.pdf. World Health Organization (WHO): Geneva. 2006. Date Accessed 9-22-2011.

32. Chaen, H. Pullulan. Food Stab., Thickeners Gelling Agents. 2010. CAPLUS AN 2010:484781(Conference; General Review).

33. Leung AY and Foster S. Encyclopedia of Common Natural Ingredients Used in Foods, Drugs, and Cosmetics. 2nd *ed*. New York: John Wiley & Sons, Inc, 1996.

34. Food and Drug Administration (FDA). 21CFR349.12. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=349.12&CFRPart=&FRSearch=. 4-1-2011. Date Accessed 10-21-2011.

35. Merck & Co., Inc. The Merck Index. http://themerckindex.cambridgesoft.com/themerckindex/Forms/Search/ContentArea/ChemBioVizSearch.aspx?FormGroupId=200000&AppName=THEMERCKINDEX&AllowFullSearch=true&KeepRecordCountSynchronized=false&SearchCriteriaId=5&SearchCriteriaValue=77-93-0&CurrentIndex=0. 2011. Date Accessed 2011.

36. Pillai R, Redmond M, and Röding J. Anit-wrinkle therapy: Significant new findings inthe non-invasive cosmetic treatment of skin wrinkles with beta-glucan. *J Cosmet Sci*. 2005;56:(May/Jume):211.

37. Svedman P, Lundin S, Hoeglund P, HammarlundC, Malmros C, and Pantzar N. Passive drug diffusion via standardized skin mini-erosion; methodological aspects and clinical findings with new device. *Pharmaceutical Research*. 1996;13:(9):1354-1359.

38. Inchem. Xanthan Gum. http://www.inchem.org/documents/jecfa/jecmono/v21je13.htm. 2010. Date Accessed 11-15-2011.

39. Lin FSD. Gellan Gum - First Draft. http://www.inchem.org/documents/jecfa/jecmono/v28je17.htm. 1991. Date Accessed 11-15-2011.

40. Mehvar R and Shepard TL. Molecular-weight-dependent pharmacokinetics of fluorescein-labeled dextrans in rats. *Journal of Pharmaceutical Sciences*. 1992;81:(9):908-912.

41. Foster BC, Gallicano KD, Whitehouse LW, McGilveray IJ, and Khan SR. Dextran sulfate disposition in the rat. *Biopharmaceutics & Drug Disposition*. 1990;11:(7):595-606.

42. Joint FAO/WHO Expert Committee on Food Additives (JEFCA). Safety Evaluation of Certain Food Additives and Contaminants. WHO Food Additive Series 44. Curdlan. http://www.inchem.org/documents/jecfa/jecmono/v44jec04.htm. 2000. Date Accessed 11-15-2011.

43. Oku T, Yamada K, and Hosoya N. Effect of pullulan and cellulose on the gastrointestinal tract of rats. *Eiyo to Shokuryo (Nutr.Diets)*. 1982;32:235-241.

44. Dixon B, Abbott PJ, Verger P, and DiNovi M. Pullulan - First draft. http://www.inchem.org/documents/jecfa/jecmono/v56je05.pdf. 2005. Date Accessed 11-9-2011.

45. Lorentsen KJ, Hendrix CW, Collins JM, Kornhauser DM., Petty BG, Klecker RW., Flexner C, Eckel RH, and Lietman PS. Dextran sulfate is poorly absorbed after oral administration. *Annals of Internal Medicine*. 1989;111:(7):561-566.

46. Yoneyama M, Okada K, Maddai T, Aga H, Sakai S, and Ichikawa T. Effects of pullulan intake in humans. *Starch Chemistry (Denpun Kagaku)*. 1990;37:(3):123-127.

47. Friberg U, Graf W, and Åberg B. Effects of prolonged dextran administration to rabbits. *Acta Pharmacol Toxicol*. 1953;9:220-234.

48. Mehvar R, Robinson MA, and Reynolds JM. Dose dependency of the kinetics of dextrans in rats: Effects of molecular weight. *Journal of Pharmaceutical Sciences*. 1995;84:(7):815-818.

49. Yamaoka T, Tabata Y, and Ikada Y. Body distribution profile of polysaccharides after intravenous administration. *Drug Delivery*. 1993;1:(1):75-82.

50. Kaneo Y, Tanaka T, Nakano T, and Yamaguchi Y. Evidence for receptor-mediated hepatic uptake of pullulan in rats. *J Control Release*. 2001;70:(3):365-373.

51. Arturson E. and Wallenius G. The intravascular persistence of dextran of different molecular sizes in normal humans. *Scandinavian Journal of Clinical and Laboratory Investigation*. 1964;16:(1):76-80.

52. Straszek SP, Adamcakova-Dodd A, Metwali N, Pedersen OF, Sigsgaard T, and Thorne PS. Acute Effect of Glucan-Spiked Office Dust on Nasal and Pulmonary Inflammation in Guinea Pigs. *Journal of Toxicology and Environmental Health, Part A*. 2007;70:(22):1923-1928.

53. Fogelmark B, Sjostrand M, Williams D, and Rylander R. Inhalation toxicity of (1→ 3)-β−D-glucan: recent advances. *Mediators of Inflammation*. 1997;6:(4):263-265.

54. Fogelmark B, Goto H, Yuasa K, Marchat B, and Rylander R. Acute pulmonary toxicity of inhaled β-1→3-glucan and endotoxin. *Agents and Actions*. 1992;35:(1-2):50-56.

55. Bonlokke JH, Stridh G, Sigsgaard T, Kjaergaard SK., Loefstedt H, Andersson K, Bonefeld-Jorgensen EC, Jayatissa MN, Bodin L, Juto JE, and Molhave L. Upper-airway inflammation in relation to dust spiked with aldehydes or glucan. *Scandinavian Journal of Work, Environment & Health*. 2006;32:(5):374-382.

56. Kim H, Cho H, Moon S, and et al. Effects of β-glucan from Aureobasidium pullulans on acute inflammation in mice. *Arch Pharm Res*. 2007;30:(3):323-328.

57. Kim KH, Chung CB, and et al. Cosmeceutical properties of levan produced by Zymomonas mobilis. *J Cosmet Sci*. 2005;56:(-):395-406.

58. Eastwood MA, Brydon WG, and Anderson DMW. The dietary effects of xanthan gum in man. *Food Additives & Contaminants*. 1987;4:(1):17-26.

59. Mitsuhashi M, Yonetama M, and Sakai S. Growth promoting agent for bacteria containing pullulan with or without dextran. 1990. (EP 0 382 355 B1):Secondary reference in INCHEM (2005).

60. Sargent EV, Adolph J, Clemmons MK, Kirk GD, Pena BM, and Fedoruk MJ. Evaluation of flu-like symptoms in workers handling xanthan gum powder. *Journal of occupational medicine.: official publication of the Industrial Medical Association*. 1990;32:(7):625-630.

61. Woodard G, Woodard MW, McNeely WH, Kovacs P, and Cronin MTI. Xanthan gum. Safety evaluation by two-year feeding studies in rats and dogs and a three-generation reproduction study in rats. *Toxicology and Applied Pharmacology*. 1973;24:(1):30-36.

62. Hirono I, Kuhara K, Yamaji T, Hosaka S, and Golberg L. Carcinogenicity of dextran sulfate sodium in relation to its molecular weight. *Cancer Letters (Shannon, Ireland)*. 1983;18:(1):29-34.

63. Tardieu D., Jaeg JP, Cadet J, Embvani E, Corpet DE, and Petit C. Dextran sulfate enhances the level of an oxidative DNA damage biomarker, 8-oxo-7,8-dihydro-2'-deoxyguanosine, in rat colonic mucosa. *Cancer Letters (Shannon, Ireland)*. 1998;134:(1):1-5.

64. Hirono I, Kuhara K, Hosaka S, Tomizawa S, and Golberg L. Induction of intestinal tumors in rats by dextran sulfate sodium. *JNCI, Journal of the National Cancer Institute*. 1981;66:(3):579-583.

65. Hirono I, Kuhara K, Yamaji T, Hosaka S, and Golberg L. Induction of colorectal squamous cell carcinomas in rats by dextran sulfate sodium. *Carcinogenesis (London)*. 1982;3:(3):353-355.

66. Tanaka T, Kohno H, Suzuki R, Yamada Y, Sugie S, and Mori H. A novel inflammation-related mouse colon carcinogenesis model induced by azoxymethane and dextran sodium sulfate. *Cancer Science*. 2003;94:(11):965-973.

67. Suzuki R, Kohno H, Sugie S, Nakagama H, and Tanaka T. Strain differences in the susceptibility to azoxymethane and dextran sodium sulfate-induced colon carcinogenesis in mice. *Carcinogenesis*. 2006;27:(1):162-169.

68. Takeuchi A, Kamiryou Y, Yamada H, Eto M, Shibata K, Haruna K, Naito S, and Yoshikai Y. Oral administration of xanthan gum enhances antitumor activity through Toll-like receptor 4. *International Immunopharmacology*. 2009;9:(13-14):1562-1567.

69. Kimura Y, Sumiyoshi M, Suzuki T, and Sakanaka M. Antitumor and antimetastatic activity of a novel water-soluble low molecular weight β-1, 3-D-glucan (branch β-1,6) isolated from Aureobasidium pullulans 1A1 strain black yeast. *Anticancer Research*. 2006;26:(-):4131-4142.

70. Yoo SH, Yoon EJ, Cha J, and Lee HG. Antitumor activity of levan polysaccharides from selected microorganisms. *International Journal of Biological Macromolecules*. 2004;34:37-41.

71. Paull, J. D. Dextrans. *Developments in biological standardization*. 1987;67:133-138.

72. Ott CM and Day DF. Modification of Natural Gums. 2000. pp.186-228. EBSCO Publishing.

73. Li H, Xu H, and et al. Enhanced welan gum production using a two-stage agitation speed control strategy in Alcaligenes sp. CGMCC2428. *Bioprocess Biosyst Eng*. 2011;34:(1):95-102.

74. Manjanna KM, Kumar TMP, and Shivakumar B. Natural polysaccharide hydrogels as novel excipients for modified drug delivery systems: A review. *International Journal of ChemTech Research*. 2010;2:(1):509-525.

75. Bresin A, Sanhajl G, and Reynaud R. Rhizobium gum: A novel cosmetic ingredient from soil to the skin. *Cosmetics & Toiletries Magazine*. 2004. 119:(4): pp.86-92.

76. Mehvar R. Dextrans for targeted and sustained delivery of therapeutic and imaging agents. *Journal of Controlled Release*. 2000;69:(1):1-25.

77. Miyake M and Kakizawa Y. Morphological study of cationic polymer-anionic surfactant complex precipitated in solution during the dilution process. *J Cosmet Sci*. 2010;61:(July/August):289-301.

78. Park SY, Bae IY, Lee S, and Lee HG. Physicochemical and Hypocholesterolemic Characterization of Oxidized Oat β-Glucan. *Journal of Agricultural and Food Chemistry*. 2009;57:(2):439-443.

79. Ding XL and Wang M. Development of a water-soluble α-carboxymethyl-β-(1,3)-glucan derived from *Saccharomyces cerevisiae*. 2008. pp.412-419.

80. Mibelle Biochemistry. 2010. 2010 CM-Glucan granulate specifications. Unpublished data submitted by Personal Care Products Council.

81. Kaplan DL, Wiley BJ, Mayer JM, and et al. Biosynthetic Polysaccharides. Chapter: 8. Shalaby SW. 1994:189-212.

82. Combie J. Properties of levan and potential medical uses. Chapter: 13. In: *Polysaccharides for Drug Delivery and Pharmaceutical Applications*. 2006:263-269.

83. Nohata Y, Azuma J, and Kurane R. Structural studies of a neutral polysaccharide produced by Alcaligenes latus. *Carbohydrate Research*. 1996;293:(2):213-222.

84. Jungbunzlauer. 2009. Safety Data Sheet: Xanthan Gum. Unpublished data submitted by Personal Care Products Council.

85. Green R. 2010. Letter from Biopolymer International to the European Commission concerning molecular weight of polymer additives. Unpublished data submitted by Personal Care Products Council.

86. Bajaj IB., Survase SA, Saudagar PS., and Singhal RS. Gellan gum: Fermentative production, downstream processing and applications. *Food Technology and Biotechnology*. 2007;45:(4):341-354.

87. Food Chemicals Codex. Rockville, MD: The United States Pharmacopeial Convention, 2008.

88. Chandrasekaran R, Radha A, and Lee EJ. Structural roles of calcium ions and side chains in welan: an X-ray study. *Carbohydrate Research*. 1994;252:(93):183-207.

89. Solabia. Active Ingredients Catalog [pamphlet]. 2011.

90.   The United States Pharmacopeia, 32nd Revision/The National Formulary, 27th edition. Rockville, MD: The United States Pharmacopeial Convention, 2009.

91. ADEKA Corporation. Technical Information. Fermented Beta-Glucan (LQ/Powder) [pamphlet]. 3-19-2009.

92. Cross GG, Jennings HJ, Whitfield DM, and et al. Immunostimulant oxidized β-glucan conjugates. *International Immunopharmacology*. 2001;1:(-):539-550.

93. Klecak J. 1998. Summary report CM-Glucan (Sodium Carboxymethyl Betaglucan) in skin care products. RCC Project 678082. Unpublished data submitted by Personal Care Products Council.

94. Hayashibara International Inc. Pullulan GRAS Notification. http://www.accessdata.fda.gov/scripts/fcn/gras_notices/215492E.PDF. 2-5-2002. Date Accessed 11-6-2011.

95. Nakamura S. Pullulan. *Journal of Synthetic Organic Chemistry, Japan*. 1984;43:(6):584-588.

96. Leathers TD. Biotechnological production and applications of pullulan. *Appl Microbiol Biotechnol*. 2003;62:(5-6):468-473.

97. Bozou JC, Gautry L, and Pianelli G. A new biopolymer for refreshment. *SOFW Journal*. 2004;130:(3):17-21.

98.   Food Chemicals Codex. 8 *ed*. Rockville, MD: United States Pharmacopeia (USP), 2012.

99.   Food Chemicals Codex. 8 *ed*. Rockville, MD: United States Pharmacopeia (USP), 2012.

100. Biopolymer International. Xanthan gum - Manufacturing Process. http://www.biopolymer-international.com/html/manufacturing.php. 2010. Date Accessed 8-31-2011.

101. Hijiya H and et al. Shaped bodies of pullulan esters and their use. 3-18-1975. 424,151:(3,871,892):Okayama, Japan:

102. Crompin JM, Garnier T, Payot T, and DeBaynast R. A new polysaccharide derived from plant rhizosphere: production, purification and physico-chemical properties. *Engineering and Manufacturing for Biotechnology*. 2002;4:(8):423-428.

103. Biopolymer International. Xanthan gum. Product description. http://www.biopolymer-international.com/html/xanthan_gum.php. 2010. Date Accessed 8-30-2011.

104. Bajaj IB., Survase SA, Saudagar PS., and Singhal RS. Gellan gum: Fermentative production, downstream processing and applications. *Food Technology and Biotechnology*. 2007;45:(4):341-354.

105. Biopolymer International. Gellan Gum - Product description. http://www.biopolymer-international.com/html/gellan_gum.php. 2010. Date Accessed 8-30-2011.

106. Booth AN, Hendrickson AP, and DeEds F. Physiologic effects of three microbial polysaccharides on rats. *Toxicology and Applied Pharmacology*. 1963;5:478-484.

107. McNeely WH and Kovacs P. The physiological effects of alginates and xanthan gum. *ACS Symp Ser*. 1975;15:269-281.

108. Babícek K, Cechova I, Simon RR, Harwood M, and Cox DJ. Toxicological assessment of a particulate yeast (1,3/1,6)-β-D-glucan in rats. *Food and Chemical Toxicology*. 2007;45:(9):1719-1730.

109. Life Science Laboratory. 1999. Single oral dose toxicity study of CM-Glucan J in mice (at a dose level of 2000 mg/kg). Test Code No. 99-IA1-1004. Unpublished data submitted by Personal Care Products Council.

110. Juntendo University, Department of Public Hygiene, School of Medicine. 1974. Report on acute toxicity test on pullulan with mice. Unpublished data reported in Hayashibara International Inc.

111. Walton KW. Investigation of the toxicity of a series of dextran sulphates of varying molecular weight. *Brit J Pharmacol*. 1954;9:1-14.

112. Williams DL, Sherwood ER, Browder IW, McNamee RB, Jones EL, and DiLuzio NR. Pre-clinical safety evaluation of soluble glucan. *International Journal of Immunopharmacology*. 1988;10:(4):405-414.

113. Robbins DJ, Moulton JE, and Booth AN. Subacute toxicity study of a microbial polysaccharide fed to dogs. *Food and Cosmetics Toxicology*. 1964;2:(5):545-550.

114. Anderson DMW, Brydon WG, and Eastwood , M. A. The dietary effects of gellan gum in humans. *Food Additives & Contaminants*. 1988;5:(3):237-249.

115. Jonker, D., Hasselwander, O., Tervilae-Wilo, A., and Tenning, P. P. 28-Day oral toxicity study in rats with high purity barley beta-glucan (Glucagel). *Food and Chemical Toxicology*. 2010;48:(1):422-428.

116. Feletti F, De, Bernardi di Valserra M, Contos S, Mattaboni, P, and Germogli R. Chronic toxicity study on a new glucan extracted from Candida albicans in rats. *Arzneimittel-Forschung*. 1992;42:(11):1363-1367.

117. Kimoto T, Shibuya T, and Shiobara S. Safety studies of a novel starch, pullulan: chronic toxicity in rats and bacterial mutagenicity. *Food and Chemical Toxicology*. 1997;35:(3/4):323-329.

118. Fogelmark B, Sjoestrand M, and Rylander R. Pulmonary inflammation induced by repeated inhalations of β¦(1→3)-D-glucan and endotoxin. *International Journal of Experimental Pathology*. 1994;75:(2):85-90.

119. Rylander R and Fogelmark B. (1→3)−β-Glucan and endotoxin modulate immune response to inhaled allergen. *Mediators of Inflammation*. 1998;7:105-110.

120. Mori H, Ohbayashi F, Hirono I, Shimada T, and Williams GM. Absence of genotoxicity of the carcinogenic sulfated polysaccharides carrageenan and dextran sulfate in mammalian DNA repair and bacterial mutagenicity assays. *Nutrition and Cancer*. 1984;6:(2):92-97.

121. Chen SN, Nan FH, Chen S, Wu JF, Lu CL, and Soni MG. Safety assessment of mushroom β-glucan: Subchronic toxicity in rodents and mutagenicity studies. *Food and Chemical Toxicology*. 2011;49:2890-2898.

122. NOTOX BV. 1996. Evaluation of the mutagenic activity of CM-Glucan in the *Salmonella lyphimurium* reverse mutation assay (with independent repeat). Notox Project 182914. Unpublished data submitted by Personal Care Products Council.

123. Life Science Laboratory. 1999. Reverse mutation study of CM-Glucan J in bacterial. Test Code No. 99-VII- 1001. Unpublished data submitted by Personal Care Products Council.

124. Magnani M, Calliari CM., de Macedo, Mori MP., de Syllos Colus I, and Castro-Gomez RJH. Optimized methodology for extraction of (1→3)(1→6)-β-D-glucan from Saccharomyces cerevisiae and in vitro evaluation of the cytotoxicity and genotoxicity of the corresponding carboxymethyl derivative. *Carbohydrate Polymers*. 2009;78:(4):658-665.

125. Ishidate M, Sofuni T, and Kishi M. Results of mutagenicity tests of fodd additives (6). *Tokishidoroji Foramu (Toxiccoogy Forum)*. 1985;8:705-708.

126. Kuroda K, Yoo YS, and Ishibashi T. Rec-assay on natural food additives. *Seikatsu Eisei*. 1989;33:15-23.

127. Ishidate M, Takizawa Y, Sakabe Y, Ishizaki M Watanabe S, Date M, and Takemoto K. Mutagenicity tests of food additives (9). *Tokishidoroji Foramu (Toxiccoogy Forum)*. 1988;11:663-669.

128. Guillot JP, Giauffret JY, Martini MC, Gonnet JF, and Soule G. Safety evaluation of gums and thickeners used in cosmetic formulations. *International Journal of Cosmetic Science*. 1982;4:(2):53-65.

129. Life Science Laboratory. 1999. Primary skin irritation study of CM-Glucan J in rabbits. Test Code No. 99-JXA4-1001. Unpublished data submitted by Personal Care Products Council.

130. Life Science Laboratory. 1999. Skin sensitization study of CM-Glucan J in guinea pigs (by maximization test method). Test Code No. 99-VIA3-1001. Unpublished data submitted by Personal Care Products Council.

131. Life Science Laboratory. 1999. Primary skin irritation test for CM-Glucan J in human subjects by closed patch test. Test Code No. 99-Xll-1010. Unpublished data submitted by Personal Care Products Council.

132. Therapy and Performance Research Institute (GTLF). 1993. Repeated patch test for skin sensitization and photoallergy of CM-Glucan (700-01) in healthy male and female volunteers. Unpublished data submitted by Personal Care Products Council.

133. Eurofins ATS. 1997. Ocular tolerance test according to NET CAM method: Sodium Carboxymethyl Beta-Glucan. Unpublished data submitted by Personal Care Products Council.

134. Liu Y, Liu J, Zhang X, Zhang R, Huang Y, and Wu C. In situ gelling Gelrite/alginate formulations as vehicles for ophthalmic drug delivery. *AAPS PharmSciTech*. 2010;11:(2):610-620.

135. Ramaiah S, Kumar TMP, and Ravi V. Studies on biopolymers for ophthalmic drug delivery. *Journal of Macromolecular Science, Part A: Pure and Applied Chemistry*. 2007;44:(2):229-234.

136. Life Science Laboratory. 1999. Primary eye irritation study of CM-Glucan J in rabbits. Test Code No.99-LXB4-1003. Unpublished data submitted by Personal Care Products Council.